Syntheses, Binding Properties, and Structures of Seven New Hemicarcerands Each Composed of Two Bowls Bridged by Three Tetramethylenedioxy Groups and a Fourth Unique Linkage^{1,2}

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Treatment of 2 mol of the bowl-shaped tetrol 1 (derived originally from resorcinol and dihydrocinnamaldehyde) with 3 mol of $TsO(CH_2)_4OTs$ gave diol 2. Eight compounds with different combinations of bridges were formed from 2 by treatment with Cs_2CO_3 and the following reagents in the presence of potential guests to give either free or complexed hemicarcerands as follows: ClCH₂Br gave 4; TsO(CH₂)₂OTs gave 5; TsO(CH₂)₃OTs gave 6; MsO(CH₂)₄OMs gave 7, a known system; $MsO(CH_2)_5OMs$ gave **8**; 2,3-bis(bromomethyl)quinoxaline gave **9**; 1,3-($ClCH_2)_2C_6H_4$ gave 10; 2,6-bis(chloromethyl)pyridine gave 11. Thirty-six fully characterized new hemicarceplexes are reported which were prepared either directly from diol 2 by the "sealing in" of the guest during introduction of the fourth bridge, or by guest exchange driven by mass law at 25 to 160 °C. The guests ranged in size from CHCl₃ to 1,2,3-(MeO)₃C₆H₃. The incarcerated guests correlated with portal sizes of their hosts. Crystal structures of $8\odot4$ -MeC₆H₄OMe and $10\odot$ CHCl₃ were determined. Changes in chemical shifts in ¹H NMR spectra of incarcerated and free guests are interpreted in terms of their locations in the hosts' inner phases. The length and nature of the unique host bridge affects the chemical shifts of the other bridges. Force field calculations of structural models for N-methylpyrrolidinone incarcerated in 4-7 were made. Approximate half-lives for decomplexation were determined for complexes involving the larger hosts and guests. Force-field calculations were made of binding energies and activation energies for decomplexations of models of $7 \odot N$ methylpyrrolidinone, $\mathbf{8} \odot N$ -methylpyrrolidinone, and $\mathbf{10} \odot N$ -methylpyrrolidinone. The activation energies for decomplexation were dissected into intrinsic and constrictive components.

Introduction

Prior publications on hemicarceplexes involved the assembly of many hosts by the four-fold bridging of two bowl-shaped cavitands such as 1 with groups, all four of which were the same. Variation in the lengths and sizes of these groups and the sizes of guests controls the propensity for complex formation and their stabilities in solution. Hemicarceplexes were prepared by guesttemplation of the shell closures, or by heating empty hosts with potential guests as solvent. A third method involves heating hosts or unstable complexes with large excesses of new guests in high-boiling solvents whose molecules are too large to enter or occupy the inner phases of hosts.³ A fourth method was described, in which 2 containing three $(CH_2)_4$ bridging groups was prepared (30-40% yields) from 1, making use of the fact that the rate of introducing the fourth bridge leading to the host is slower than those rates for the first three. The fourth bridge was introduced in the presence of large excesses of guests that, in certain cases, were too big or unstable to provide complexes made by the other three methods.⁴ For example, 7 O(CH₂CH₂)₂NCHO was prepared from 2 and MsO(CH₂)₄OMs in (Me₂N)₃PO (HMPA) containing O(CH₂CH₂)₂NCHO. This "sealing in of guest"

method of complexation had the additional synthetic advantage that the fourth bridge could be the same (leading to 7^4) or different than the first three, as in 4-6and 8-11. In this manner 3 was prepared, which was found to complex potassium picrate, the picrate ion being incarcerated but the K⁺ being ligated by the six oxygens of the fourth bridge.⁴

We report here the syntheses and study of new host systems **4–6** and **8–11** from diol **2** and appropriate dihalides, ditosylates, or dimesylates in the presence of Cs₂CO₃ and potential guests.

Results and Discussion

Syntheses. Diol 2 was prepared in 40% yield from 1 and 3 equiv of $MsO(CH_2)_4OMs$ in a mixture of Cs_2CO_3 and NMP⁵ at 25 °C for 18 h.³ This material served as starting material for the syntheses of the seven new host systems in which the fourth bridge (R of 4-6, 8-11) differs from the three $(CH_2)_4$ bridges of 2. Chart 1 summarizes the reagents, reaction conditions, solvents, product structures, and yields for these shell closures. The four aprotic dipolar solvents used were NMP,5 DMSO, DMA,⁵ and HMPA. Bridge donor groups and products were as follows: CH2BrCl led to 4ONMP, **4** \odot DMSO, and **4** \odot DMA; TsOCH₂CH₂OTs gave **5** \odot NMP, 5ODMSO, and 5ODMA; TsO(CH₂)₃OTs provided 6ONMP, 6ODMSO, 6ODMA, and empty 6 (HMPA as solvent); MsO(CH₂)₄OMs gave 7⁶ (HMPA as solvent); MsO(CH₂)₅-

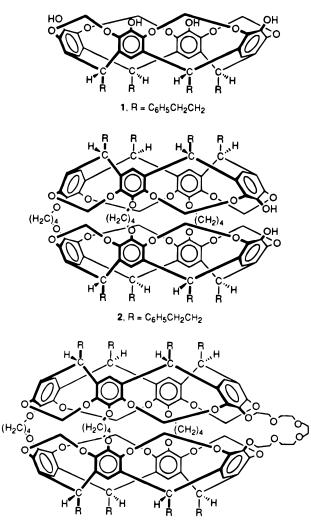
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⁽²⁾ We warmly thank the U. S. Public Health Service for supporting grant GM-12640, and Dr. Kurt Loening for assistance with nomenclature.

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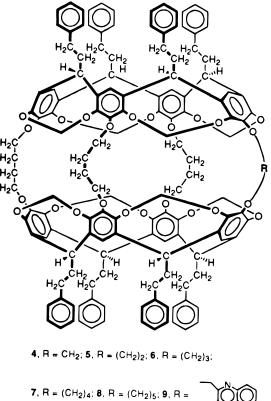
⁽⁵⁾ NMP stands for *N*-methylpyrrolidinone; DMA for $(CH_3)_{2^-}$ NCOCH₃. GB/SA is the Generalized Born radii (GB)/solvent-accessible surface area (SA), an empirical solvation model. It is a semianalytical treatment of solvation and provides a volume-based continuum model for the electrostatic (polarization) component. $^{\rm 13}$



3, $R = C_6H_5CH_2CH_2$

OMs in NMP probably initially produced 80 NMP, whose NMP was replaced with CHCl₃ to give **8**OCHCl₃ (stable to manipulation) during chromatographic purification with CHCl₃ as the mobile phase; 2,3-bis(bromomethyl)quinoxaline provided 90NMP, 90DMSO, and 901,4- $(MeO)_2C_6H_4$ (HMPA as solvent containing a 100:1 molar ratio of guest $1,4-(MeO)_2C_6H_4$ to **2**). This last reaction, carried out at 24-50 °C, exemplifies the "sealing in" of a relatively large guest during introduction of the fourth bridge. With m-xylyl dichloride in NMP, whatever **10**ONMP was produced initially went to **10**OCHCl₃ during isolation. Similarly, 2,6-bis(chloromethyl)pyridine ultimately gave **11** OCHCl₃ although the shell closure was conducted in NMP. The temperatures for the shell closures varied from 24-75 °C, and the times from 48-53 h.

Chart 2 indicates the starting materials and products for the thermally induced formations of new hemicarceplexes by either guest exchanges, or from empty host plus guests. Where possible, the new guest served as solvent for the starting empty host or complex. When necessary for solubility reasons Ph_2O or HMPA were used as solvents whose molecular volumes are too large to occupy the inner phases of any of these hosts. In such cases, guest-to-host molar ratios of at least 100 were employed. Temperatures and times for these equilibrations with one exception ranged respectively from 140-165 °C, with times of 72-96 h. The exception was when 7 and naphthalene dissolved in Ph_2O were heated at 200 °C



7, $R = (CH_2)_4$; 8, $R = (CH_2)_5$; 9, R =

for 2400 h, 7 \odot naphthalene was formed in 40% yield. In all the experiments of Chart 2, the cooled reaction mixtures were flooded with methanol which precipitated the complexes, most of which were purified by thick layer chromatography with CHCl₃ as the mobile phase. Complexes $9\odot1,4-(MeO)_2C_6H_4$, $10\odot4-MeC_6H_4OMe$, $10\odot1,4-(MeO)_2C_6H_4$, and $11\odot4-MeC_6H_4OMe$ were used without chromatographic purification. With the exception of $7\odot$ naphthalene, in which equilibrium was probably not established, the yields varied from 80-92%.

Isolated Products Correlate with Portal Size. Comparisons of CPK models suggest the inner volumes of these hosts differ very little from one another, but that the two portals that flank the last bridge introduced increase in size and conformational adaptivity in passing from 7 to 8, and from 7 to 10 or 11. The portals flanked by two (CH₂)₄ groups already present in 2 are not greatly affected by the length of the fourth bridge. These two portals are invariant and composed of 26-membered rings, as compared to the other two portals that flank the fourth bridge. The latter vary in ring size with changes in the fourth bridge as follows: 4, 23-membered rings; 5, 24-membered rings; 6, 25-membered rings; 7, 26-membered rings; 8, 27-membered rings; 9, 26-membered rings with four bridging atoms coplanar; 10, 27membered rings with five of the bridging atoms coplanar; and 11, 27-membered rings with five of the bridging atoms coplanar. To enter or exit the inner phase of these hosts, potential guests must pass through these rings, and for the guests studied, the larger portals are going to be the most used. With 4-7 and 9, these are likely to be the original 26-membered portals; with 8, 10, and 11, the larger portals are probably used.

Chart 1. Fourth Bridge Donors React with Diol 2 To Give Either Host⊙Guest Complexes or Hosts When Heated in the Presence of Cs₂CO₃

		Ċ	`N-Mə (NMP) /	ı		Mə S ↓	Me (DMSC))		Me N	-C, (DM/	A)		Me Me	Me N Me N − N (HMPA) ^a V Me	ı
bridge-	Condi	tions	Product		Cond	itions	Product		Cond	litions	Product		Cond	itions	Product	
donor to 2	T (*)	t(h)	kind	yld(%)	T (°)	1(h)	kind	yld(%)	T (*)	t(h)	kind	yld(%)	T (*)	t(h)	kind	yld(%)
CH2BrCl	65	48	40NMP	53	65	48	40DMSO	51	65	48	4 ODMA	54				
TsOCH ₂ CH ₂ OTs	75	48	50NMP	45	75	48	5 ODMSO	41	75	48	5 ODMA	42				
TsO(CH ₂)3OTs	75	48	60NMP	64	75	48	6 ODMSO	60	75	48	6 ODMA	64	75	48	6	46
MsO(CH ₂) ₄ OMs ^b	25-60	72	7 Onmp	55									25	48	7	60
MsO(CH ₂)5OMs	25-50	53	8 OCHCl3c	79												
	24-50	53	90nmp	78	24-50	53	90dmso	72					24-5() 53	9 ⊙1,4-(MeO)2C6H4 ^d	¹ 34
	25-50	53	100CHCl3c	81												
	25-50	53	110CHCl3°	77												

reaction media and source of guests

^a CPK examinations show that HMPA is too large to exist in the inner phases of these hosts.

^b Taken from reference (4). From the same reference, 2 + MsO(CH₂)₄OMs in O(CH₂CH₂)₂NCHO-Cs₂CO₃ at 25 ^cC for 48 h gave 70O(CH₂CH₂)₂NCHO (85%).

^c During chromatographic isolation with CHCl₃, host ONMP exchanges guest to give host OCHCl₃, driven by mass law.

^d HMPA which served as solvent contained a 100:1 molar ratio of 1,4-(McO)₂C₆H₄:2.

This analysis grossly correlates with observations incidental to the syntheses of the hosts. Thus the originally formed "sealed in" complexes of Chart 1 involving 4-7 and 9 survived their isolation in the presence of CHCl₃ during chromatography. Even empty 7 survived isolation without complexing CHCl₃.⁴ However, 8, 10, and 11 lost their putative original guest (NMP) during isolation in favor of complexing CHCl₃. Models show little constrictive hindrance to the departure of NMP and entrance of CHCl₃ to the interior of these hosts. On the other hand, the slow complexation of naphthalene by empty 7 (2400 h to produce 40% complex at 200 °C) correlates with the fact that in CPK models complexation requires considerable modification of stable conformations and some bond angle adjustments in many parts of the host.

The numbers of non-hydrogen atoms in the guests introduced into hosts in this study ranges between 4 (CHCl₃, (CH₃)₂SO) and 12 (1,2,3-(MeO)₃C₆H₃), the most common guests containing 8 to 10 non-hydrogen atoms, six often in the form of a benzene ring. Host 7 in other studies was found to incarcerate 33 other guests of widely differing structures possessing most of the common functional groups, none of which contained more than 10 non-hydrogen atoms.^{6,7} Those selected for the current study were chosen generally for their size, solubility properties, boiling points, simplicity of their ¹H NMR spectra, and probability of forming a hemicarceplex. Almost all organic compounds containing 8 or less nonhydrogen atoms are good candidate guests for the hosts of this study. However, it is highly probable that hosts **8**, **10**, and **11** are capable of incarcerating larger guests than **4**–**7** and **9**, although the only example reported here is **10** \odot 1,2,3-(MeO)₃C₆H₃.

Crystal Structures of 804-MeC6H4OMe and 10⊙CHCl₃. Crystal structures of 8⊙4-MeC₆H₄OMe⊍ 2PhNO₂ (R = 0.162) and 10 \odot CHCl₃ \cup 2PhNO₂·2Ph-NO₂ (R = 0.17) were determined.⁸ Chart 3 provides both side and partial top stereoviews of the complexes. In the latter, the two sets of four oxygens that terminate the intermolecular bridges are connected by heavy lines to form two near planar squares attached to one another by the four interhemispheric bridges, the rest of the host being omitted. These top view partial structures both demonstrate that the carbons and hydrogens of the four bridges all lie outside the volume of a solid figure defined by the eight oxygens at its corners. This geometry reflects the fact that the unshared electron pairs of all eight oxygens face inward toward the cavity, and their bonds to the bridges must diverge from the cavity. This arrangement gives an out-in-out orientation of unshared electron pairs for the three oxygens attached to adjacent carbons on each of the eight benzene rings of the host, which provides for the greatest compensation of dipoles and lowest energy.

Crystalline $8\odot4-MeC_6H_4OMe\cup2PhNO_2$ is isostructural with more than 25 hemicarceplexes of $7\odot$ guest \cup

⁽⁶⁾ Robbins, T. A.; Knobler, C. B.; Bellew, D. R.; Cram, D. J. J. Am. Chem. Soc. **1994**, *116*, 111–122.

⁽⁷⁾ Robbins, T. A.; Cram, D. J. J. Am. Chem. Soc. **1993**, 115, 12199–12200.

⁽⁸⁾ The authors have deposited atomic coordinates for these structures with the Cambridge Crystallographic Data Center. The coordinates can be obtained, on request, from the Director, Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge, CB2 1EZ, UK.

Chart 2.	New Hemicarceplexes Prepared by Mass-Law Driven Guest Exchange or from Free
]	Host-Complexing Solvent or Solutes, Both Methods at Elevated Temperature

	medium conditions		itions	product		
starting material	solvent	<u>solute</u>	T (°C)	t (h)	kind	yield (%)
40DMA	1,4-(Me) ₂ C ₆ H ₄		140	72	401,4-(Me) ₂ C ₆ H ₄	85
50DMA	1,4-(Me) ₂ C ₆ H ₄		140	72	$501,4-(Me)_2C_6H_4$	88
6	1,4-(Me) ₂ C ₆ H ₄		140	72	$601,4-(Me)_2C_6H_4$	9 0
7	Ph ₂ O ^a	©0	200	2400	70 00	40 ^b
7 °	C ₆ H ₅ OCH ₂ CH=CH ₂		160	96	7OC ₆ H ₅ OCH ₂ CH=CH ₂	9 0
8	CHCl ₂ CHCl ₂		140	72	80CHCl ₂ CHCl ₂	92
80CHCl3	Ph ₂ O ^a	ÔÔ	165	72	80 00	84
80CHCl3	Ph ₂ O ^a	1,4-(MeO) ₂ C ₆ H ₄	165	72	801,4-(MeO) ₂ C ₆ H ₄	92
80CHCl3	1,2-(MeO) ₂ C ₆ H ₄		165	72	801,2-(MeO) ₂ C ₆ H ₄	86
80CHCl3	1,3-(MeO) ₂ C ₆ H ₄		165	72	801,3-(MeO) ₂ C ₆ H ₄	89
80CHCl3	4-MeOC ₆ H ₄ Me		165	72	804-MeOC ₆ H ₄ Me	92
80CHCl3	2-HOC ₆ H ₄ Me		165	72	802-HOC ₆ H ₄ Me	9 0
10OCHCl3	CHCl ₂ CHCl ₂		140	72	10OCHCl ₂ CHCl ₂	91
10OCHCl3	Ph ₂ O ^a	ÔÔ	165	72	100 🔘	85
10OCHCl ₃	1,2-(MeO) ₂ C ₆ H ₄		165	72	1001,2-(MeO) ₂ C ₆ H ₄	87
10OCHCl3	1,3-(MeO) ₂ C ₆ H ₄		165	72	1001,3-(MeO) ₂ C ₆ H ₄	89
10OCHCl3	Ph ₂ O ^a	1,4-(MeO) ₂ C ₆ H ₄	165	72	$1001,4-(MeO)_2C_6H_4$	90
10OCHCl3	4-MeOC ₆ H ₄ Me		165	72	1004-MeOC ₆ H ₄ Me	90
10OCHCl3	Ph ₂ O ^a	1,2,3-(MeO) ₃ C ₆ H ₃	165	72	1001,2,3-(MeO) ₃ C ₆ H ₃	80
110CHCl ₃	Ph ₂ O ^a	ÔÔ	165	72	110 🔘	87
110CHCl3	1,2-(MeO) ₂ C ₆ H ₄		165	72	1101,2-(MeO) ₂ C ₆ H ₄	84
11OCHCl3	1,3-(MeO) ₂ C ₆ H ₄		165	72	1101,3-(MeO) ₂ C ₆ H ₄	86
110CHCl3	4-MeOC ₆ H ₄ Me		165	72	1104-MeOC ₆ H ₄ Me	90

^a (Me₂N)₃PO (HMPA) and Ph₂O are each too large to occupy the interiors of these hosts. ^bThe authors thank Siavash Kurdistani for this result. ^c Taken from reference (4).

2PhNO₂ containing four [O(CH₂)₄O] bridges, and therefore the parameters for the two cavitand moieties and for the two PhNO₂ molecules could be used to locate the guest, the three $O(CH_2)_4O$ bridges, and the $O(CH_2)_5O$ bridge. Because the complex is crystallographically centrosymmetric, the guest is required to be disordered. A "pair" of bridges is also required to have disorder (the unlike bridges related by the center of symmetry). The 4-MeC₆H₄OMe guest molecule in the host cavity is disordered because it lies on the crystallographic center of symmetry and is further disordered because it lies on either diagonal of the rectangular solid described by the eight terminal oxygens of the bridges as seen in the top partial stereoview. The four bridge-terminating oxygens attached to each bowl are coplanar within 0.01 (2) Å, and the planes are 4.10 Å distant from one another. These two planes are not rotated around the polar axis with respect to one another (see top view). The two PhNO₂ molecules are associated with the CH₂CH₂Ph "feet" attached to the northern and southern hemispheres, with the nitro groups turned toward the globe of the host (see Chart 3, side view).

In the crystal structure of 10 OCHCl₃ U2PhNO₂. $2PhNO_2$, the complex lies on a two-fold axis. The CHCl₃ is located in the cavity, but is disordered. Each set of four CH₂CH₂Ph groups surrounds a PhNO₂ molecule, with the nitro group turned toward the globe. The other two $PhNO_2$ molecules are interstitial. The $PhNO_2$ molecules are omitted from the drawings of Chart 3. The two sets of four oxygens that terminate the four interhemispheric bridges form two planes (\pm 0.02 (2) Å). The two planes miss being parallel to each other by 5.7°. The oxygen atoms of each interhemispheric bridge are distant from one another by the values (Å): $OCH_2C_6H_4CH_2O_4$ 4.47 (2); O(CH₂)₄O (flanking bridges), 4.02 (2) and 4.06 (2); $O(CH_2)_4O$ (opposite bridge), 3.57 (2). The average distance between the two oxygen planes is 4.03 Å, less than the 4.10 Å distance in $8\odot4$ -MeC₆H₄OMe. The carbons and hydrogens of these four bridges all lie outside the solid defined by the eight apical oxygens. The two

 Chart 3. Stereoviews of Crystal Structures of Hemicarceplexes

 side views

 top partial views

 Image: Colspan="2">Image: Colspan="2">Image: Colspan="2" Image: Colspan="2" Image:

10OCHCl3

oxygen planes are slightly rotated and displaced from their best central axis with respect to one another. The top view of **10** shows that the two portals flanking the $OCH_2C_6H_4CH_2O$ bridge are slightly more open than the two portals flanking the $O(CH_2)_5O$ bridge in **8**. The long bridges are clearly visible at about 1 o'clock and 5 o'clock in the top views of **8** and **10**, respectively.

Correlations between Guest Structures and Changes in Chemical Shifts of Incarcerated and Free Host Protons in ¹H NMR Spectra. Table 1 lists δ for 12 guests dissolved in CDCl₃, 43 hemicarceplexes of these guests dissolved in CDCl₃, and $\Delta \delta$ values for the guest protons ($\Delta \delta = \delta_{\rm free} - \delta_{\rm complexed}$). In all cases except for the OH protons in 7 \odot 2-MeC₆H₄OH and **8** \odot 2-Me-C₆H₄OH the δ values moved upfield upon incarceration, due to the shielding effects of the faces of the eight aryl groups that line much of the surface of the inner phase. The $\Delta \delta$ values range from a low of -0.76 (in **8** \odot 2-Me-C₆H₄OH) to a high of 4.41 ppm (in 7 \odot 1,3-(MeO)₂C₆H₄).

Model examinations of 40NMP, 50NMP, 60NMP, **7**ONMP, and **9**ONMP indicate that the Me of the guest must occupy one of the polar, bowl-shaped interiors of the four hosts, and thus anchor the guest in the four hosts. The other protons of the CH₂CH₂CH₂ parts of the guest are arranged in an arc which stretches like the handle of a tea kettle through the equator and past the other hemisphere of the inner phase to attach to the C=O (the spout of the tea kettle) whose axis points roughly toward the lower parts of the bridging groups of the hosts. Thus the $\Delta \delta$ values of the four kinds of protons provide a sort of map of the shielding character of the different parts of the inner surfaces of the closely related hosts. The guest is presumed to rotate rapidly on the ¹H NMR time scale around the axis of the N-Me bond, which is roughly coincident with the long polar axis of the host.

100CHCl3

It also seems likely that $\Delta \delta$ values should measure how far the Me protons are thrust into the shielding faces of the four aryls. Table 1 indicates the $\Delta\delta$ values for the five N-Me protons decrease in the following order with increases in the effective lengths of the unique bridge of the host as follows: $CH_2C=CCH_2$ (9), 3.87 ppm; CH_2 (4), 3.83; CH₂CH₂ (5), 3.63; CH₂CH₂CH₂ (6), 3.60; CH₂CH₂- CH_2CH_2 (7), 3.59 ppm. By effective lengths, we mean the effect on the long axis of the inner phase of the hosts. Interestingly, the coplanarity requirement of the fouratom 2,3-dimethylenequinoxaline bridge of 9 forces it in CPK models to rotate about its C_2 axis and thus act as a bridge even slightly shorter than the CH₂ bridge of 4. The effective lengths of the other four bridges fall in the expected order, with little difference between the (CH₂)₃ and (CH₂)₄ bridges. The H^b protons provide a similar sequence: CH₂C=CCH₂, 3.19; CH₂, 3.13; CH₂CH₂, 2.95; CH₂CH₂CH₂, 2.83; CH₂CH₂CH₂CH₂, 2.82 ppm. The H^c protons give a somewhat different order: CH₂, 3.15; CH₂C=CCH₂, 3.05; CH₂CH₂, 2.79; (CH₂)₃, 2.72; and (CH₂)₄, 2.68 ppm. The only distinguishable H^d protons were when the host bridges were $CH_2C=CCH_2$ and CH_2 , which gave the same $\Delta \delta$ values (1.46 ppm). As expected, for each complex, the three or four different protons available provided decreases in $\Delta \delta$ values in the order $H^a > H^b \approx H^c > H^d$. In models, the H^c and H^d protons cannot penetrate as deeply into the polar regions of the hosts as can the H^a protons of the Me group.

The $\Delta\delta$ values for Me₂SO (singlets) are smaller than those for H^a of NMP and vary much less with changes in the host's unique bridges, probably because this guest is smaller than NMP and less closely held. The order of $\Delta\delta$ with changes in the unique bridges is as follows: CH₂C=CCH₂, 3.17; CH₂, 3.14; (CH₂)₃, 2.97; (CH₂)₄, 2.95; CH₂CH₂, 2.94 ppm. Again, the *effective lengths* are the

Table 1. Effect of Host Structure on 500 MHz ¹H NMR Spectra of Guests in CDCl₃ at 25 °C

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Guest	proton	free	40gue		50gues		60gu		70gu		9⊙gue	st
Structure	proton	guest δ	δ	Δδ	δ	Δδ	δ	Δδ	δ	Δδ	δ	Δδ
ÇH3ª	Hª	2.70 (s)	-1.13 (s)	3.83	-0.93 (s)	3.63	-0.90 (s)	3.60	-0.89 (s)	3.59	-1.17 (s)	3.87
H ^o	Нр	2.23 (t)	-0.90 (t)	3.13	-0.72 (t)	2.95	-0.60 (l)	2.83	-0.59 (t)		-0.96 (t)	3.19
	Hc	1.90 (q)	-1.25 (q)	3.15	-0.89 (q)	2.79	-0.82 (q)	2.72	-0.78 (q)	2.68	-1.15 (q)	3.05
H ^c NMP H ^d	Hq	3.26 (t)	1.80 (t)	1.46	hidden		hidden		hidden		1.80 (t)	1.46
11111												
Å												
S	Hª	2.46 (s)	-0.68 (s)	3.14	-0.48 (s)	2.94	-0.51 (s)	2.97	-0.49 (s)	2.95	-0.71 (s)	3.17
DMSO	1.10	0.09 (-)	1.00 (-)	4 00	1 (7 ())		1 (0 ()					
	Hp Hp		-1.92 (s) -0.90 (s)	4.00 3.92	-1.67 (s) -0.56 (s)	3.75 3.58	-1.08 (s) -0.53 (s)		-1.64 (s) -0.42 (s)			
H ₃ C ^C N ^{CH₃b}	H¢	2.94 (s)	-0.90 (s) 1.58 (s)	1.36	-0.38 (s) 1.64 (s)	3.38 1.30	hidden	3.33	-0.42 (s) 1.61 (s)			
DMA CH3°	11.	2.74 (3)	1.50 (3)	1.50	1.04 (8)	1.50	Inducti		1.01 (3)	1.55		
CH₃ª		0.00 (-)	0.00.(-)		102 ()		A A 1 (-)	4.99	0.00 (.)	4 40		
	Нр На		-2.02 (s)	4.34	-1.93 (s)	4.25	-2.01 (s)	4.33	-2.08 (s)			
\checkmark	H	7.07 (s)	5.85 (s)	1.22	5.97 (s)	1.10	5.94 (s)	1.13	5.90 (s)	1.77		
CH3												
			7⊙gue		80gues	_	9⊙gue		10⊙g	-	110gu	
			δ	Δδ	δ	Δδ	δ	Δδ	δ	Δδ	δ	Δδ
CI CI H-C-C-H*												
H-Ċ-Ċ-Hª CI CI	Ha	5.95 (s)	4.75 (s)	1.20	4.29 (s)	1.66			4.35 (s)	1.60		
Hª	Ha	7.66 (d)	hiddon		hidden				hidden		hidden	
H ^b	Нр			4 21	3.36(br s)	3 04			3.35(br s)	3 05		2 02
		7.50 (u)	5.07(01 3)	4.41	2.50(01 3)	5.74			5.55(01 3)	3.75	5.57 (m)	5.75
QCH₃ª												
H ^b	Hª	3 77 (s)	-0.46 (s)	4.23	-0.41 (s)	4.18	-0.45 (s)	4.22	-0.38 (s)	4 15		
	Нр		5.82 (s)	1.03	5.84 (s)	1.01	5.81 (s)	1.04	5.84 (s)			
осн₃		0.02 (0)	0.02 (0)		0.0. (5)			••••				
OCH ₃ *	T 10	2 80 (-)	0 (1 (-)		0.55 (-)	4.25			0.52 (.)	4 9 9	0 60 (-)	4.20
H ^b	Ha Hp	5.80 (s) 6.50 (s)	-0.61 (s) 5.42 (s)	4.41 1.08	-0.55 (s) 5.48 (s)	4.35 1.02			-0.53 (s) 5.48 (s)		-0.50 (s) 5.49(br s)	
	Hc	6.54 (d)	3.42 (s) 4.82 (d)	1.72	5.02 (d)	1.52			5.03 (d)		5.00 (d)	1.54
H¢ OCH3	Hq	7.20 (t)	4.02 (u) 4.12 (t)	3.08	hidden	1.52			hidden	1.51	hidden	1.54
H ^c ỌCH₃ ª				5.00								
	Ha	3.88 (s)	hidden		hidden	• •			hidden		hidden	
	Hp	6.9 (m)	hidden	1.0	5.46(br s)	1.4			5.45(br s)	1.5	5.55(br s)	1.3
Hc	Hc	6.9 (m)	4.98(br s)		hidden				hidden		hidden	
OCH3*	Ha	3.78 (s)	-0.35 (s)	4.13	-0.28 (s)	4.06			-0.26 (s)		-0.29 (s)	4.07
	Нр	6.81 (d) 7.09 (d)	5.87 (s) 5.87 (s)	0.94	5.84 (d)	0.97			5.87 (d) 6.01 (d)		5.78 (d) 5.90 (d)	1.03
H°	Hq Hq	2.29 (s)	-2.11 (s)	1.22 4.40	5.95 (d) -1.98 (s)	1.14 4.27			-1.96 (s)		-1.98 (s)	1.19 4.27
ĊH₃⁴	11-	2.29 (3)			-1.70 (3)				-1.90 (3)	4.2.5	-1.90 (3)	4.27
	Ha	5.45 (s)	5.91 (s)	-0.46	6.21 (s)	-0.76						
	Нр	2.31 (s)	-1.75 (s)	4.06	-1.68 (s)	3.99						
H ^e H ^c	Hc	6.82 (d)			hidden							
Hd	Hq	6.92 (t)	6.13 (t)	0.79	6.14 (t)	0.78						
	He Lif	7.14 (t)	3.05 (t)	4.09	3.22 (t)	3.92						
OCH3ª	Hf H ^a	7.18 (d) 3.85 (s)	5.73 (d)	1.45	5.98 (d)	1.20			-0.41 (s)	4 26		
	Hp Hp	3.85 (s) 3.86 (s)							-0.41 (s) 3.05 (s)			
	Hc	6.58 (d)							hidden	0.01		
H ^d OCH ₃ *	Hq	6.99 (d)							hidden			
est for the CH_C=			[bridge	, but	the	CU.	the Ua on	dцb,		f DM	A incorco	ratad ir

shortest for the CH₂C=CCH₂ and CH₂ bridges, but the other three $\Delta \delta$ values do not vary significantly with the numbers of methylenes.

As observed for the carceplex of $MeCONMe_2$ (DMA) whose host was identical to **4** except all four bridges were

 CH_2 , the H^a and H^b methyls of DMA incarcerated in **4–7** give substantially higher $\Delta\delta$ values (range 4.00–3.44 ppm) than the H^c methyls (range, 1.36–1.30 ppm). The H^a and H^b methyls lie on the long axis of DMA, which must be close to being coincident with the long axes of

the host cavities. Thus the H^a and H^b methyls are located in the two polar (most shielding) parts of the host's interior, and the H^c methyls lie in the equatorial region of the bridges. The H^a protons' $\Delta\delta$ value is 4.00 with the shortest unique bridge (CH₂), but decreases to 3.75, 3.76, and 3.72 ppm, respectively, as that bridge becomes CH₂-CH₂, (CH₂)₃, and (CH₂)₄. The H^b protons' $\Delta\delta$ values decrease more markedly and regularly with the lengthening of the unique bridge as follows: CH₂, 3.92; CH₂-CH₂, 3.58; (CH₂)₃, 3.55; (CH₂)₄, 3.44 ppm. The equatorially-located H^c protons provide $\Delta\delta$ values that vary little and irregularly with bridge length.

In models, 1,4-Me₂C₆H₄ fits comfortably in 4-7 with each of the two methyl groups occupying one of the two hemispheric bowls with the long axes of host and guest being essentially coincident. The H^a protons of these methyls exhibit $\Delta \delta$ values that change relatively little with the unique bridge changes as follows: CH₂, 4.34; CH₂CH₂, 4.25; (CH₂)₃, 4.33; (CH₂)₄, 4.40 ppm. The nonmonatonic orders in some of these patterns probably reflect different degrees of rotation of the two hemispheres with respect to one another as the unique bridge lengths change, and the host adapts to the steric requirements of these relatively rigid guests. The hosts more than the guests have bond angle and conformational degrees of freedom that can vary cumulatively to minimize the energies of the complexes of 4-7 and 9. The enlargement of the host's portal should be more temperature-sensitive than the compressibility of the guest.

Both host inner phases and guest volumes are larger for the complexes whose $\Delta \delta$ values are reported in the bottom two-thirds of Table 1. The two protons (H^a) of CHCl₂CHCl₂ in models occupy niches defined by the much larger Cl atoms, and therefore cannot reach the inner shielding surfaces of the host when incarcerated. Accordingly, the $\Delta \delta$ values for **7**, **8**, and **10** with unique bridges (CH₂)₄, (CH₂)₅, and 1,3-(CH₂)₂C₆H₄ are 1.20, 1.66, and 1.60 ppm. In contrast, with naphthalene as guest, the protons are exposed, and in models of their complexes, the H^b protons rest against the high-shielding faces of the northern and southern hemispheres of the host. Accordingly, the $\Delta \delta$ values for H^b are high and are as follows (unique bridges identify the host): $(CH_2)_4$, 4.21; (CH₂)₅, 3.94; 1,3-(CH₂)₂C₆H₄, 3.95; 2,6-(CH₂)₂pyridine, 3.93 ppm. As expected, the shortest unique bridge, $(CH_2)_4$, provides the highest $\Delta \delta$ value.

The next three guests of Table 1 are the three isomeric dimethoxybenzenes. The most complementary relationship between host and guest with regard to the partners sharing the largest surface area is found in the complexes of 7, 8, 9, and 10 with 1,4-(MeO)₂C₆H₄. The H^a methyl protons in models of the complexes are thrust nonsymmetrically into each of the two hemispheres of the hosts. Their $\Delta \delta$ values are as follows: unique bridge (CH₂)₄, 4.23; (CH₂)₅, 4.18; CH₂C=CCH₂, 4.22; and 1,3-(CH₂)₂C₆H₄, 4.15 ppm. The inner phases of these complexes appear to be very similar to one another in shape. More surprisingly, the same thing is observed for the 1,3- $(MeO)_2C_6H_4$ isomer. In models, the two methyls nicely fit into the two hemispheres of the host, which is consistent with the high $\Delta \delta$ values for the H^a protons of the two methyls, as follows: $(CH_2)_4$, 4.41; $(CH_2)_5$, 4.35; 1,3-(CH₂)₂C₆H₄, 4.33; and 2,6-(CH₂)₂pyridine, 4.30 ppm. In models, the H^b proton is located in the equatorial region of the four complexes, and their $\Delta \delta$ values range from 1.08 to 1.01 ppm. The H^c protons' $\Delta \delta$ values are somewhat higher, since they contact the edge of the aryl faces, and their $\Delta\delta$ values show the greatest range, 1.51 to 1.72 ppm. It is more difficult in models of complexes of 1,2-(MeO)_2C_6H_4 to locate the contacting parts since each OMe group inhibits its neighboring OMe group from penetrating the polar parts of the cavity. In none of the ¹H NMR spectra of the four complexes that were taken could the δ values for the H^a methyl protons be determined because of the overlap of host and guest signals. Probably a family of structures, nonequilibrating on the NMR time scale and close in energy, is responsible for the uninformative spectra.

Like 1,4-(MeO)₂C₆H₄ as guest, 4-MeOC₆H₄Me is highly complementary in shape to the inner phases of these larger hosts in molecular models. The $\Delta\delta$ values for the four different kinds of protons in 4-MeOC₆H₄Me in its four complexes are very close to one another: for H^a, with (CH₂)₄ as a fourth bridge, 4.13; (CH₂)₅, 4.06; 1,3-(CH₂)₂C₆H₄, 4.04; 2,6-(CH₂)₂pyridine, 4.07 ppm. The respective $\Delta\delta$ values for H^b, H^c, and H^d in the four complexes are the following: 0.94, 0.97, 0.94, and 1.03; 1.22, 1.14, 1.08, and 1.19; 4.40, 4.27, 4.25, and 4.27 ppm.

Both model examination and $\Delta \delta$ values for the H^b methyl protons of 2-HOC₆H₄Me indicate the Me group occupies one of the polar regions of the host in the two complexes prepared and examined. The $\Delta \delta$ values are as follows: unique bridge (CH₂)₄, 4.06; (CH₂)₅, 3.99 ppm. The phenolic hydroxyl protons (H^a) are actually deshielded, with $\Delta \delta$ values of -0.46 and -0.76, respectively. These acidic protons probably hydrogen bond the oxygens that terminate the bridges at that end of the host in each complex whose hemispheres are occupied by the Me groups. As expected, the H^e aryl protons para to the anchoring Me groups of the guests provide high $\Delta \delta$ values of 4.09 for the (CH₂)₄ bridged and 3.92 for the (CH₂)₅ bridged complexes, respectively. The remaining aryl protons with observable $\delta,\ H^d$ and $H^f,$ in the two complexes have $\Delta \delta$ values between 0.78 and 1.45 ppm.

The most crowded complex prepared is $10 \odot 1,2,3$ -(MeO)₃C₆H₃, whose unique bridge is 1,3-(CH₂)₂C₆H₄. Models of this complex resemble those of $10 \odot 1,3$ -(MeO)₂C₆H₄. The aryl plane of this guest is tilted with respect to the equatorial plane of the host, with one H^a methyl protruding into one polar region of the host, and the second H^a methyl protruding into the other polar region. In models of $10 \odot 1,2,3$ -(MeO)₃C₆H₃, the middle MeO group is close to being coplanar with its attached aryl group, which places the H^b methyl protons in the low-shielding equatorial region. The $\Delta\delta$ values are consistent with such a structure, being 4.26 for H^a and 0.81 for H^b in $10 \odot 1,2,3$ -(MeO)₃C₆H₃, the former being comparable to 4.33 ppm for H^a in $10 \odot 1,3$ -(MeO)₂C₆H₄.

Important conclusions emerge from these correlations between models of host–guest complexes, and the $\Delta\delta$ shielding–deshielding patterns of incarcerated guests. (1) When only a single CPK model of a complex can be assembled in which the various parts of each complexing partner are pretty well fixed with respect to one another, the correlation between structural conclusions based on ¹H NMR $\Delta\delta$ values and the model structure is high. Examples involve complexes of DMA, 1,4-(Me)₂C₆H₄, 1,4-(MeO)₂C₆H₄, 1,2,3-(MeO)₃C₆H₃, and 4-MeOC₆H₄Me. (2) When several models of a given host complex can be assembled, the ¹H NMR spectral peaks become broader, and the structural information contained in $\Delta\delta$ values becomes more restricted and ambiguous. Examples are complexes of 1,2-(MeO)₂C₆H₄ and Me₂SO.

Representative Examples of Effects on Host NMR Spectra as Lengths and Character of One Bridge of 7 and Its Guests are Changed. Figure 1 records the interesting parts of the 500 MHz ¹H NMR spectra in CDCl₃ at 25 °C of the host parts of complexes involving **4**–**6** and **9**. Replacement of one $O(CH_2)_4O$ bridge in **7** by one O–A–O bridge in **4**–**6** and **9** destroys the formal C₄ axis of **7**, but leaves a formal mirror plane defined by the two oxygens that terminate the unique bridge, and the two oxygens that terminate the most remote $O(CH_2)_4O$ bridge. This leads in principle to three different kinds of *interhemispheric* OCH_2 protons, four different kinds of *intrahemispheric* OCH_2 protons, and two different methine C₃CH protons.

Some of these closely related protons provide resolved signals in the spectra of Figure 1. For example, in the spectrum of 40NMP, the three different H^e signals for OCH2eCH2CH2CH2eO are close together, whereas in $4\odot 1,4$ -(Me)₂C₆H₄, they are widely separated, probably because of their varying locations with respect to the shielding magnetic fields of their tightly-held arvl guest. In contrast, two sets of H^b and H^d signals are visible in the $4\odot$ NMP spectrum but only one set (H^b and H^d) in that of $4\odot 1,4$ -(Me)₂C₆H₄. In the spectrum of $5\odot$ NMP, two H^e, two H^d, but only one H^b signals are visible. In the spectrum of 6ONMP, one H^e, one H^b, and two H^d peaks are apparent. In all of these four spectra only one methine signal is observed, but in those of **9**ONMP and $9 \odot 1,4$ -(MeO)₂C₆H₄, two are clearly discernible. The spectrum of **9**ONMP contains a single H^b, two H^d signals, but only one H^e peak. In contrast, that of 901,4-(MeO)₂C₆H₄ provides all three H^e, two H^b, and two H^d signals. These examples as well as many others found in the Experimental Section show how extensively the ¹H NMR spectra of the host protons vary with guest character.

Force Field Calculations of Structural Models for 4 ONMP-7 ONMP Complexes. The conformations of hosts and complexes were explored with force field calculations using AMBER* in the program MACRO-MODEL.⁹ This force field has proven useful for the investigation of hemicarceplex conformations and the dynamic processes involved in complexation and decomplexation.¹⁰⁻¹² For computational simplification, the eight $CH_2CH_2C_6H_5$ groups of 4-7 were replaced with CH_3 groups to provide 4-Me, 5-Me, 6-Me, and 7-Me, respectively. Conformations of the four interhemispheric bridges of 4-Me to 7-Me were explored with a Monte Carlo (MC) search by varying the torsional angles. Host 7-Me was constructed by the graphical input module in MACRO-MODEL. The starting geometries for 6-Me, 5-Me, and 4-Me were obtained by removing one, two, and three methylenes, respectively, from one of the interhemispheric bridges of 7-Me, and then optimizing the resulting structures with the AMBER* force field. The total number of MC steps used for each structure was 500, and each MC step began with the geometry of the leastused structure of the previous MC steps. Many lowenergy conformers of 7-Me were located. The conformation of 7-Me derived from the crystal structure of 701,4-

(9) Mohamadi, F.; Richards, N. G. J.; Guida, W. C.; Liskamp, R.; Caufield, C.; Chang, G.; Hendrikson, T.; Still, W. C. *J. Comput. Chem.* **1990**, *11*, 440.

- host proton labels:
- H^a, OCH2^aO, OCH2^aCH2^aO or OCH2^aCH2CH2^aO
- H^b, outer proton of OCH₂^bO, intrahemispheric
- H^C, methine
- H^d , inner proton of OC H_2^d O, intrahemispheric
- H^e, OCH₂^eCH₂CH₂CH₂^eO
- H^f, guest H
- H^g, (CH₂^g)₂quinoxaline of 9

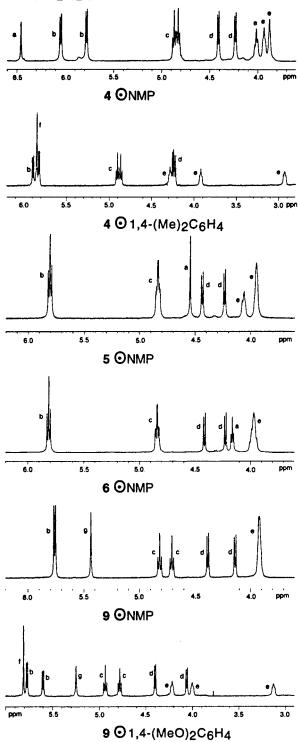


Figure 1. Partial 500 MHz ¹H NMR spectra of hosts of **4**⊙Guest, **5**⊙Guest, **6**⊙Guest, and **9**⊙Guest.

 $Me_2C_6H_4^6$ is very close structurally to the global minimum obtained by the MC search, and its energy is only 0.6

⁽¹⁰⁾ Sheu, C.; Houk, K. N. J. Am. Chem. Soc. **1996**, 118, 8056–8070.

⁽¹¹⁾ Nakamura, K.; Houk, K. N. *J. Am. Chem. Soc.* **1995**, *117*, 1853–1854.

⁽¹²⁾ Houk, K. N.; Nakamura, K.; Sheu, C.; Keating, A. E. Science 1996, 273, 627–629.



Figure 2. Superposition of 10 low-energy conformers of 7-Me.

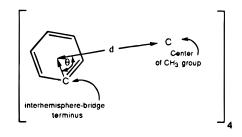
kcal mol⁻¹ higher by AMBER*. The northern and southern bowls of the hemicarcerand are very rigid and the flexibility of the molecule is mainly due to the interhemispheric bridges. Figure 2 is the superposition of 10 low-energy conformers, all of which are within 1.5 kcal mol⁻¹ of the global minimum.

Starting geometries for the NMP complexes were obtained manually by docking the NMP guest inside the cavity of the hemicarcerand in various orientations. These were energy-minimized with the AMBER* force field. The lowest-energy conformer of each carceplex was then used as the starting geometry in an optimization utilizing the simulated annealing method. The cooling process was linear and continuous from 500 K to 50 K over a 1000 ps molecular dynamics simulation. The structure of the lowest-energy conformer for each complex is found in Figure 3.

The best plane of the guest in the calculated structures of Figure 3 is roughly placed in the plane of the page, with the *N*-methyl pointing upward into the north polar cap. In all four structures, the best plane of the guest and page roughly bisects two of the portals, with two of the bridges flanking the C=O group, and two bridges flanking the two H's of the NCH₂ group. The four $CH_2CH_2CH_2N$ hydrogens of the guest point generally toward the southern polar part of the host, which is consistent with the substantial upfield shifts in their ¹H NMR spectra.

A general and important feature of these calculated structures is that the electron pairs of the oxygen termini of the *interhemispheric bridges* all face inward, opposite to the directions in which the *intrahemispheric spanner* oxygens face, thus minimizing the energy of their dipole–dipole interactions. Consequently, the carbon chains of the interhemispheric bridges lie outside of the distorted cube defined by the eight oxygens to which these chains are attached. In these respects the calculated structure of host 7 in 7-Me \odot NMP resembles the crystal structures of 7 \odot Guests, many of which have been determined,⁶ and also the structures of 8 \odot 4-MeC₆H₄OMe and 10 \odot CHCl₃ reported here (Chart 3).

Analysis of the calculated structures of **4-Me** to **7-Me** provides the average distance between two planes, each defined by the four aryl carbons at the termini of the four interhemispheric bridges in the respective northern and southern hemispheres. Table 2 lists the results. These distances (Å) increase monotonically with increasing lengths of the unique bridges in the hosts and range from 5.1 to 6.0 Å. Two other parameters locate the center of the guest NMP methyl group with respect to the center of the four proximate benzenes of the host: the distance (*d*) between these two centers: the average deviation from 90° of the angle θ between vector **d** and one connecting the center of the benzene with the terminus of the interhemispheric bridge. These parameters are visualized in diagram 1.

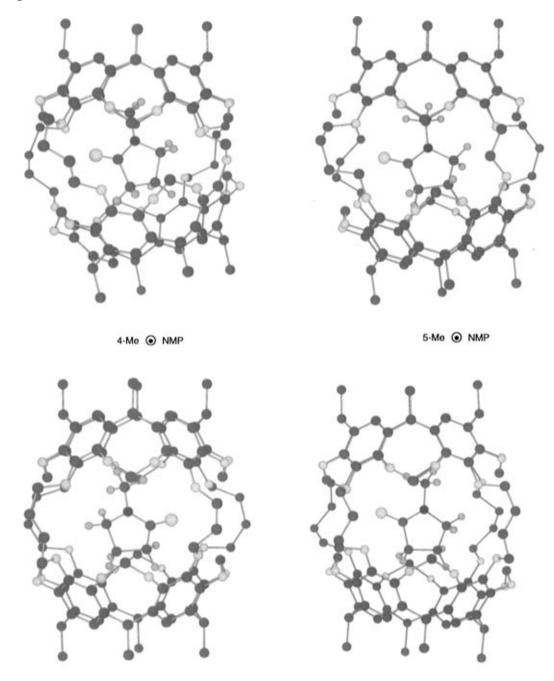


These distances and θ angle deviations are listed in Table 2, and they correlate, respectively, with the number of methylenes in the unique bridges as follows: CH₂, 3.5 Å and 9.8°; (CH₂)₂, 3.8 Å and 4.9°; (CH₂)₃, 4.0 Å and 4.5°; (CH₂)₄, 3.9 Å and 4.4°. These parameters also correlate with the upfield chemical shift changes in the ¹H NMR spectral $\Delta\delta$ values for the methyl group of NMP upon incarceration in **4**–**7**, listed in Table 2.

Approximate Half-Lives for Decomplexation of Complexes Involving Largest Hosts and Guests. Much qualitative information dealing with complexes stable or unstable to isolation conditions is found in Chart 1. Isolation involved evaporation of aprotic dipolar solvents under vacuum under 45 °C, flooding the reaction mixtures with methanol, and subjecting the methanolwashed precipitate to thick layer chromatography with CHCl₃ as the mobile phase. Complexes of **4**, **5**, **6**, **7**,⁶ and 9 with NMP, DMSO, and DMA as guests are stable to these manipulations, whereas free 6 and 7 did not complex under similar conditions (Chart 1). In contrast, 80NMP, 100NMP, and 110NMP went to 80CHCl₃, **10** \odot CHCl₃ and **11** \odot CHCl₃ under the same conditions. Complexation of **4**–**7** or **9** and decomplexation of their complexes involves guests passing through 23- to 26membered rings. Only guests as small as CH₂Cl₂ or pentane (but not CHCl₃) readily entered and departed the interior of **7** in solution at 25 °C.⁶ However, CHCl₃ readily passed through the 27-membered ring portals of 8, 10, and 11. Temperatures of 100-200 °C appear needed for hosts with 26-membered and 27-membered ring portals to complex or decomplex guests containing 6-12 atoms other than hydrogen (Chart 2 and prior work^{3,6}).

Because little was known about the kinetic stability of complexes of hosts with 27-membered ring portals, we determined the approximate half-lives for decomplexation in CDCl₃ at 25 °C of the complexes between **7**, **8**, **10** and 4-MeC₆H₄OMe, 4-MeOC₆H₄OMe, and 3-MeOC₆H₄OMe. Table 3 records the $t_{1/2}$ values for those decomplexations that could be detected.

Complexes of 7 with all three guests gave no ^{1}H NMRdetectable uncomplexed guest after 336–720 h. Half-



6-Me
NMP

7-Me
NMP

Figure 3. Force field calculated structures derived for molecules of $4 \odot NMP$, $5 \odot NMP$, $6 \odot NMP$, and $7 \odot NMP$, simplified by substitution of CH₃ for CH₂CH₂C₆H₅ groups in the hosts to give, respectively, **4-Me** \odot NMP, **5-Me** \odot NMP, **6-Me** \odot NMP, and **7-Me** \odot NMP. In the drawings, H atoms are omitted from the hosts and the unique bridges are on the right, toward the viewer.

Table 2. Results of Force Field Calculations of Structures of NMP Complexes of Hosts 4-Me-7-Me^a

		average distances b	etween centers (Å)		
complex modeled	unique bridge	interhemispheric bridge termini ^b	NMP methyl to aryl faces ^c	average deviation of Θ from 90 ° ^c	observed ¹ H NMR $\Delta \delta$ for guest CH_3 (ppm) ^d
4-MeONMP	OCH ₂ O	5.1	3.5	9.8	3.83
5-Me⊙NMP	$O(CH_2)_2O$	5.7	3.8	4.9	3.63
6- Me ⊙NMP	O(CH ₂) ₃ O	5.9	4.0	4.5	3.60
7-Me ⊙NMP	$O(CH_2)_4O$	6.0	3.9	4.4	3.59

^{*a*} The eight $CH_2C_6H_5$ feet of hosts **4**–**7** have been replaced by CH_3 feet to simplify the calculations. ^{*b*} Hemispheric planes defined by aryl carbon termini of bridges. ^{*c*} See Diagram 1. ^{*d*} H^a protons' $\Delta \delta$ values, Table 1.

life values for decomplexations of $8\odot4$ -MeC₆H₄OMe, $8\odot3$ -MeOC₆H₄OMe, and $8\odot4$ -MeOC₆H₄OMe are as follows: >720 h, >600 h, and 310 h. Half-life values for decomplexations of $10\odot3$ -MeOC₆H₄OMe, $10\odot4$ -MeC₆H₄OMe,

and 10°·4-MeOC₆H₄OMe decreased respectively as follows: 250 h, 85 h, and 8 h.

These results emphasize how large a difference one methylene can make in a portal's ability to allow or

Table 3. Decomplexation Half-Lives in Hours in CDCl₃ at 25 $^\circ\text{C}$

		host		
guest	no.	unique	largest	t _{1/2} (h)
structure		bridge	ring (portal)	
сн,	7	O(CH ₂) ₄ O	26-membered	>> 720ª
\bigcirc	8	O(CH ₂) ₅ O	27-membered	>720 ^b
H ₃ C [*]	10	1,3-O(CH ₂) ₂ C ₆ H ₄	27-membered	85
° ^{⊂ ۲}	7	O(CH ₂) ₄ O	26-membered	>> 336ª
\bigcirc	8	O(CH ₂) ₅ O	27-membered	310
н₃с-0	10	1,3-O(CH ₂) ₂ C ₆ H ₄	27-membered	8
сн ₃	7	O(CH ₂) ₄ O	26-membered	>> 720ª
U	8	O(CH ₂) ₅ O	27-membered	>600 ^b
H₃C ⁻⁰	10	1,3-(OCH ₂) ₂ C ₆ H ₄	27-membered	250

^a No decomplexation detected after this time. ^b Less than 10% decomplexation after this time.

disallow passage of a guest from the inner to the bulk phase of a host at 25 °C. Thus 7^oguest⁶ and 12^oguest¹⁴ complexes containing 26-membered ring portals are stable indefinitely at 25 °C in solvent (such as CDCl₂-CDCl₂) with guests as small as CH₃CH₂I, CH₃COCH₃, (CH₂)₄O, and CH₃CO₂CH₂CH₃,^{3,6} as well as larger guests such as 4-MeC₆H₄OMe, 3-MeOC₆H₄OMe, and 4-MeOC₆H₄-OMe in CDCl₃. Host 8 Oguest with two 27-membered ring portals has decomplexation half-lives in the hundreds of hours, and **10** guest with two 27-membered ring portals with a *m*-xylyl group substituted for a $(CH_2)_3$ moiety in the middle of the unique bridge has half lives that range from 250 to 8 h for the three disubstituted benzene guests. Thus the *m*-xylyl group offers somewhat less resistance to guest passage than the $(CH_2)_3$ group, probably because of the latter's six hydrogens.

Force Field Calculations of Binding Energies and Activation Energies for Decomplexation of 7-Me \odot NMP, 8-Me \odot NMP, and 10-Me \odot NMP. To obtain further insight regarding the differences in stabilities of complexes 7 \odot NMP, 8 \odot NMP, and 10 \odot NMP, binding energies in the gas phase and in chloroform were calculated using the AMBER* force field program mentioned previously. As before, the calculations were simplified by replacing the eight CH₂CH₂C₆H₅ groups of each host with eight CH₃ groups to give, respectively, 7-Me \odot NMP, 8-Me \odot NMP, and 10-Me \odot NMP. The binding energies are defined by the expression

$$\Delta E = E_{\text{(complex)}} - [E_{\text{(host)}} + E_{\text{(guest)}}]$$

and are listed in Table 4 for both the gas phase and in $CHCl_3$ solution (with GB/SA^5 chloroform treatment).

Gas phase calculations were also performed to estimate the activation energies for guest escape following a procedure described earlier.^{10,11} A reaction coordinate, λ , was defined along which the guest was forced to pass through the larger of the equatorial portals.¹¹ As illustrated in Figure 4, the dotted lines connect a dummy atom (Du) with the four aryl carbon atoms 20 Å distant attached to the two interhemispheric bridges that define

Table 4. Force Field AMBER* Calculations of Binding Energies and Activation Energies for Decomplexation of 7-Me^ONMP, 8-Me^ONMP, and 10-Me^ONMP, and the Dissection of the Activation Energies for Decomplexation into Intrinsic and Constrictive Components

		-		
	energies		complex	
phase	(kcal mol ⁻¹)	7-Me ⊙NMP	8-MeONMP	10-MeONMP
gas	ΔE	-19.3	-19.8	-18.3
ČHCl₃	ΔE	-11.5	-13.4	-12.2
gas	$\Delta E^{*}_{(ext{decomplex})}$	39.0	27.1	21.3
gas	$\Delta E^{\dagger}_{(intrinsic)}$	19.3	19.8	18.3
gas	$\Delta E^{\dagger}_{\text{(constrictive)}}$	19.7	7.3	3.0

one of the larger portals. The reaction coordinate connects Du with the center of the methyl of incarcerated NMP. By gradually decreasing the distance between the guest molecule and the defined dummy atom, the activation energy for the guest escape process was estimated by energy minimization for each step. For the host molecules with all their intrahemispheric bridges (spanners) in their favored chair conformations, the activation energy barriers for the decomplexation of NMP from model hosts **7-Me**, **8-Me**, and **10-Me** were calculated. Figure 5 is a plot of energy versus distance for the three complexes from which the $\Delta E^{+}_{(decomplex)}$ values were taken. The absolute values listed (Table 4) of 39.0, 27.1, and

The absolute values listed (Table 4) of 39.0, 27.1, and 21.3 kcal mol⁻¹ are probably somewhat high, but their relative values are meaningful. Particularly interesting is the dissection of these activation energies into intrinsic and constrictive components, using the equation,

$$\Delta E^{\dagger}_{\text{(constrictive)}} = \Delta E^{\dagger}_{\text{(decomplex)}} - (-\Delta E) = \Delta E^{\dagger}_{\text{(complexation)}}^{14}$$

Table 4 lists the $\Delta E^{\dagger}_{\text{(constrictive)}}$ values, which are as follows: for **7-Me** \odot NMP, 19.7; for **8-Me** \odot NMP, 7.3; for **10-Me** \odot NMP, 3.0 kcal mol⁻¹. The percent contribution made by constrictive binding to the total activation energy for the three respective decomplexations is 51%, 27%, and 14%.

The thermodynamic ΔE values found in Table 4 correlate well with expectations based on molecular model examinations.

The ΔE values for the calculated binding energies for **7-Me** \odot NMP, **8-Me** \odot NMP, and **10-Me** \odot NMP in the gas phase provide an average of 19.1 \pm 0.8 kcal mol⁻¹, and in CHCl₃, an average of 12.4 \pm 1.0 kcal mol⁻¹. The maximum spread in each set of three values is only 2 kcal mol⁻¹, which correlates with the facts that the NMP guest is common to the three complexes, the three hosts differ only in one of their bridges, and these unique bridges differ mainly in their lengths and in the small differences in numbers of stabilizing host–guest contacts available in these hosts.

The gas phase transition state energies for complexation-decomplexation differ dramatically, that for **7-Me** \odot NMP being \approx 12 kcal mol⁻¹ higher than that for **8-Me** \odot NMP, which in turn is 5.8 kcal mol⁻¹ higher than that for **10-Me** \odot NMP. This order correlates with the order listed in Table 3 for the $t_{1/2}$ values for the decomplexations of the same hosts as follows: **7** \odot guests > **8** \odot guests > **10** \odot guests. Thus the differences between the kinetic stabilities of the three hemicarceplexes are much greater than the differences in thermodynamic stabilities.

Striking features of the energy-distance profile for decomplexation of the three model hosts shown in Figure

⁽¹³⁾ Still, W. C.; Tempczyk, A.; Hawley, R. C.; Hendrickson, T. J. Am. Chem. Soc. 1990, 112, 6127–6129.

⁽¹⁴⁾ Cram, D. J.; Blanda, M. T.; Paek, K.; Knobler, C. B. J. Am. Chem. Soc. **1992**, 114, 7765–7773.

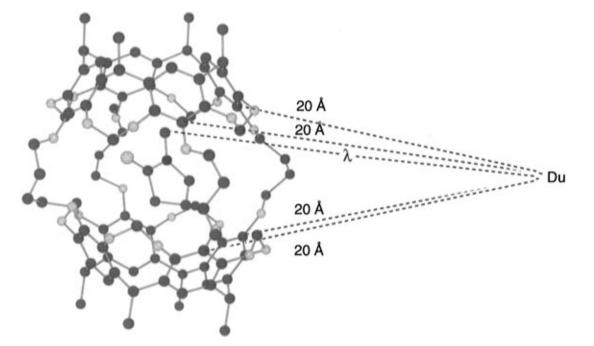


Figure 4. Definition of reaction coordinate λ for activation energy calculations. See text.

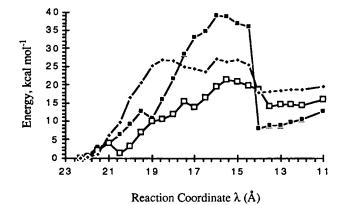


Figure 5. Energy profile for the decomplexation of the NMP guest in vacuum. ■: **7-Me**⊙NMP; **◆**: **8-Me**⊙NMP; □, **10-Me**⊙NMP.

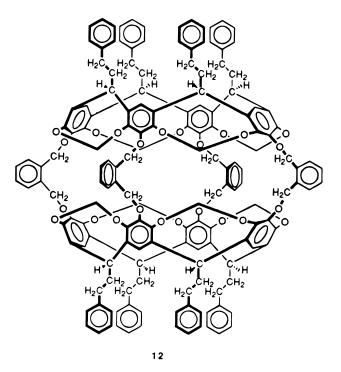
5 are as follows: (1) the energy falls very abruptly at a distance of about 14 Å, particularly for **7-Me** \odot NMP; (2) the curves contain some fine structure and several maxima; (3) the energy peaks spread over 5–6 Å for all three complexes. All three features correlate with what is suggested by the mechanical force and deformations needed to separate host from guest in CPK models of these complexes. Large numbers of bond angles must be adjusted, particularly in the host, to synchronize with guest rotations and with guest penetrations and expansions of the portals. The distance the guest must travel before it is free of the expanded portal in CPK models is in the 5–7 Å range. Even multiple mechanical barriers are encountered in these model separations.

Although hosts with longer fourth bridges ((CH₂)₅ and 1,3-(CH₂)₂C₆H₄) than (CH₂)₄ do not have significantly greater inner volumes (as seen in Table 4, the stabilization energies for **7-Me** \odot NMP, **8-Me** \odot NMP, and **10-Me** \odot NMP are very similar), the two portals that flank the longer fourth bridges dramatically increase in size and conformational adaptivity for guest passages. The portal effect can be seen in Figure 5, where the energy peak for **7-Me** \odot NMP (26-membered ring portals) is somewhat sharper and narrower than the peaks for 27-

membered ring portals (8-MeONMP, 10-MeONMP), which spread over 6-7 Å. In earlier studies, 10-12 we proposed an alternative mechanism involving the interconversions of the intrahemispheric spanners of hemicarcerands from chairlike to boatlike conformations to explain the escape pathway of acetonitrile molecules from the tetrasulfide hemicarceplex.11 This gating phenomenon also plays an important role in obtaining stable complexes.¹² The conversion of the intrahemispheric spanners from low-energy chairlike to higher-energy boatlike conformations enlarges the portals and reduces the steric repulsion between host and guest. This is especially true when the interhemispheric bridges are short (for example, 20-membered ring portals). When the interhemispheric bridges are longer, the methylene gate plays a less important role. The size and shape of the guest molecule also affect the importance of the gating mechanism. In the present case, the steric repulsive energy reduced by the gating process is approximately equal to the energy cost for the host to adopt the higherenergy conformation.

The greatest weaknesses of the activation energy calculations are the omissions of solvent and entropy. In an earlier study, the decomplexation rate for 12 ODMA¹⁴ was found to vary by a factor of about 50 as solvent was changed from C₆D₅Br to C₆D₅CD₃. Free energies, enthalpies, and entropies of activation for decomplexation of 12ODMA, 12OMeCO2CH2Me, 12OMeCOCH2Me, and **12** \odot MeC₆H₅ in 1,2-(CD₃)₂C₆D₄ were measured at 100 °C. The contributions of ΔS^{\ddagger} to the ΔG^{\ddagger} for the four complexes varied from 25 to 51%. The contribution of constrictive binding to the free energies of activation for decomplexations of the four complexes varied from 82 to 88%. Host 12 has four 26-membered ring portals as does 7, but each OCH₂C₆H₄CH₂O bridge in **12** is conformationally less adaptive than each $O(CH_2)_4O$ bridge in 7. Clearly, much exploration of decomplexation phenomena remains to be done.

Summary. Seven new carcerand or hemicarcerand systems have been synthesized, all of which contain two rigid bowls connected by three O(CH₂)₄O bridges put in



place by a shell closure. In a separate step, a fourth unique bridge was introduced by sealing into the host's inner phase appropriately sized guests in the medium. Three of the unique bridges are shorter (OCH₂O, $O(CH_2)_2O$, and $O(CH_2)_3O$, one is about the same length $(2,3-(OCH_2)_2$ quinoxaline), and three are longer $(O(CH_2)_5O)$, $1,3-(OCH_2)_2C_6H_4$, and $2,6-(OCH_2)_2$ pyridine) than the other three bridges. Thirty-six new hemicarceplexes, involving thirteen different guests, were prepared either by sealing in or by thermally-induced guest exchange driven by mass law. They were characterized, and their ¹H NMR spectral changes were correlated with changes in the fourth bridge. The crystal structures of two complexes were determined and compared with expectations based on molecular model examinations. Force field AMBER* calculations were made of the structures of four NMP complexes containing homologously related unique bridges (O(CH₂)_nO, n = 1-4) and were similar to those based on crystal structures of similar hemicarceplexes. Half-lives for decomplexations of hemicarceplexes involving three of the largest hosts and guests were correlated with portal macroring sizes. Decomplexations were modeled with AMBER* force field calculations and the results found to qualitatively correlate with experimental facts.

Experimental Section

General. All chemicals were reagent grade and used directly unless otherwise noted. Dimethylacetamide (DMA), *N*-methylpyrrolidinone (NMP), and dimethyl sulfoxide (DMSO) were stored over (24 h heated at 320 °C) 3-Å molecular sieves and degassed under high vacuum just before use. A 360-MHz spectrometer was used to record ¹H NMR spectra unless otherwise noted. Spectra taken in CDCl₃ were referenced to residual CHCl₃ at 7.26 ppm. FAB MS were determined on a ZAB SE instrument with 3-nitrobenzyl alcohol (NOBA) as a matrix and Xe as carrier gas. Gravity chromatography was performed on silica gel 60 (70–230 mesh). Thin-layer chromatography involved glass-backed plates (silica gel 60, F₂₄₅, 0.25 mm).

8,9,10,11,39,40,41,42-Octahydro-1,18,26,28,53,55,63,74octaphenethyl-34,47-(epoxybutanoxy)-20,24:57,61-dimethano-2,52:3,51:16,30:17,29-tetrametheno-1*H*,18*H*,26*H*,

28H,53H,55H-bis[1,3]benzodioxocino[9,8-d:9',8'-d']bis-[1,3]benzodioxocino[9',10':17,18;10",9":25,26][1,3,6,11,14, 16,19,24]octaoxacyclohexacosino[4,5-j:13,12-j]bis[1,3]benzodioxocin-65,72-diol, Stereoisomer 2. Into a 1 L oneneck round-bottom flask equipped with a magnetic stirrer and blanketed with argon were placed 1 g (1.0 mmol) of 1, 200 mL of degassed NMP, and 6.5 g of Cs_2CO_3 . The reaction mixture was stirred at 25 °C for 1 min after which 0.97 g (4.0 mmol) of 1.4-butanediol dimesylate was added. The mixture was stirred for 16-18 h and then poured into 500 mL of 10% NaCl (aq). After 30 min, the precipitate that formed was filtered and chromatographed with 0.5% EtOAc in CH₂Cl₂ followed by 3% EtOAc in CH₂Cl₂ to give 2 (30–40%): ¹H NMR δ 1.98 (4 H, br s); 2.03 (8 H, br s); 2.48 (16 H, m); 2.68 (16 H, m); 3.88 (4 H, br s); 3.92 (8 H, br s); 4.26 (8 H, overlapping d, *J* = 8.1 Hz); 4.82 (8 H, m); 5.85 (4 H, d, J = 7.0 Hz); 5.97 (4 H, d, J = 6.9Hz); 6.21 (2 H, s); 6.64 (2 H, s); 6.82 (4 H, s); 6.85 (2 H, s); 7.16 (16 H, m); 7.23 (24 H, m); FAB MS m/e (2194.9, M⁺), 2197.1 (100). Anal. Calcd for C140H130O24 5H2O: C, 73.54; H, 6.17. Found: C, 73.67; H, 6.04.

8,9,10,11,39,40,41,42-Octahydro-1,18,26,28,53,55,63,77octaphenethyl-34,47-(epoxybutanoxy)-20,24:57,61-dimethano-2,52:17,29-dimetheno-3,51,16,30-(methynoxymethanoxymethyno)-1H,18H,26H,28H,53H,55H-bis[1,3]benzodioxocino[9,8-d:9',8'-d]bis[1,3]benzodioxocino[9', 10':17,18;10",9":25,26][1,3,6,11,14,16,19,24]octaoxacyclohexacosino[4,5-j:13,12-j']bis[1,3]benzodioxocin, Stereoisomer 4 ONMP. Procedure A. Diol host 2⁴ (100 mg, 0.045 mmol), 50 mL of NMP, 1 g of pulverized Cs₂CO₃, and 30 μ L (0.46 mmol) of BrCH₂Cl were stirred at 65 °C for 24 h, and 30 µL (0.46 mmol) of BrCH₂Cl was added. After stirring at 65 °C for another 24 h, the solvent was removed in vacuo and the residue dissolved in $\ensuremath{\mathsf{CHCl}}_3.$ The remaining solids were filtered through a 1 cm pad of Celite, and the solvent was rotary evaporated to ~3 mL volume and poured into 100 mL of methanol. The precipitate that formed was filtered and chromatographed on a preparative TLC plate with CHCl₃ to give 55 mg (53%) of $4 \odot$ NMP: ¹H NMR δ –1.25 (2 H, q); –1.13 (3 H, s); –0.90 (2 H, t, J = 7.2 Hz); 1.80 (2 H, t, J = 7.0 Hz); 1.95 (4 H, br s); 1.97 (4 H, br s); 2.17 (4 H, br s); 2.49 (16 H, m); 2.67 (16 H, m); 3.89 (4 H, br s); 3.94 (4 H, t); 4.01 (4 H, t, J = 7.1 Hz); 4.24 (4 H, d, J = 7.1 Hz); 4.42 (4 H, d, J = 7.1Hz); 4.86 (8 H, m); 5.79 (4H, d, J = 7.3 Hz); 6.05 (4H, d, J = 7.3 Hz); 6.46 (2 H, s); 6.84 (6 H, s); 6.89 (2 H, s); 7.15 (16 H, m); 7.23 (24 H, m); FAB MS, m/e (2306.0, M⁺), 2307.0 (100). Anal. Calcd for C₁₄₆H₁₃₉NO₂₅: C, 75.99; H, 6.07. Found: C, 76.23; H, 5.75.

4⊙**DMSO.** Application of procedure A to 100 mg (0.045 mmol) of diol host **2**, 50 mL of DMSO, 1 g of Cs₂CO₃, and 60 μ L (0.92 mmol) of BrCH₂Cl gave 52 mg (51%) of **4**⊙DMSO: ¹H NMR δ −0.68 (6 H, s); 1.95−2.19 (12 H, m); 2.49 (16 H, m); 2.68 (16 H, m); 3.82 (4 H, t, J = 7.1 Hz); 3.91 (4 H, br s); 3.97 (4 H, t, J = 7.1 Hz); 4.13 (4 H, d, J = 7.3 Hz); 4.32 (4 H, d, J = 7.3 Hz); 4.86 (8 H, m); 5.85 (4H, d, J = 7.3 Hz); 6.09 (4H, d, J = 7.3 Hz); 6.54 (2 H, s); 6.80 (2 H, s); 6.84 (4 H, s); 6.92 (2 H, s); 7.17 (16 H, m); 7.24 (24 H, m); FAB MS *m*/*e* (2284.9, M⁺), 2287.1 (40); 2209 (100). Anal. Calcd for C₁₄₃H₁₃₆O₂₅S: C, 75.11; H, 5.99. Found: C, 75.36; H, 5.98.

4⊙**DMA.** Application of procedure A to 100 mg (0.045 mmol) of diol host **2**, 50 mL of DMA, 1 g of Cs₂CO₃, and 60 μ L (0.92 mmol) of BrCH₂Cl gave 56 mg of **4**⊙DMA (54%): ¹H NMR δ −1.92 (3 H, s); −0.90 (3 H, s); 1.58 (3 H, s); 1.85−2.22 (12 H, m); 2.49 (16 H, m); 2.68 (16 H, m); 3.82 (4 H, t, J = 6.9 Hz); 3.92 (4 H, br s); 4.00 (4 H, t, J = 6.9 Hz); 4.16 (4 H, d, J = 7.2 Hz); 4.54 (4 H, d, J = 7.2 Hz); 6.47 (2 H, s); 6.77 (2 H, s); 6.83 (4 H, s); 6.92 (2 H, s); 7.17 (16 H, m); 7.24 (24 H, m); FAB MS m/e (2294.0, M⁺) 2295.3 (30), 2208.3 (100). Anal. Calcd for C₁₄₅H₁₃₉NO₂₅: C, 75.86; H, 6.10. Found: C, 76.09; H, 6.37.

4 \odot **1**,**4**-**Me**₂**C**₆**H**₄. **Procedure B.** Into a Pyrex test tube capped with a rubber septum were placed 30 mg (0.013 mmol) of **4** \odot DMA and 1 mL of *p*-xylene. This mixture was heated at 140 °C for 3 days and poured into 50 mL of methanol. The precipitate that formed was filtered and chromatographed on a preparative TLC plate with CHCl₃ to give 26 mg (85%) of **4** \odot 1,4-Me₂C₆H₄: ¹H NMR δ –2.02 (6 H, s); 1.09 (4 H, m); 2.08

(4 H, m); 2.17 (4 H, m); 2.52 (16 H, m); 2.69 (16 H, m); 2.93 (4 H, t); 3.92 (4 H, br s); 4.24 (8 H, t, J = 7.8 Hz); 4.27 (4 H, t); 4.89 (8 H, m); 5.82 (4H, d, J = 6.9 Hz); 5.85 (4 H, s); 5.89 (4H, d, J = 6.9 Hz); 6.82 (2 H, s); 6.84 (2 H, s); 6.94 (2 H, s); 7.16 (16 H, m); 7.23 (24 H, m); FAB MS, m/e (2313.0, M⁺) 2314.0 (40), 2208.3 (100). Anal. Calcd for C₁₄₉H₁₄₀O₂₄·2H₂O: C, 76.13; H, 6.17. Found: C, 76.14; H, 5.99.

8,9,10,11,39,40,41,42-Octahydro-1,18,26,28,53,55,63,78octaphenethyl-34,47-(epoxybutanoxy)-20,24:57,61-dimethano-2,52:17,29-dimetheno-3,51,16,30-(methynoxyethanoxymethyno)-1H,18H,26H,28H,53H,55H-bis[1,3]benzodioxocino[9,8-d:9',8'-d']bis[1,3]benzodioxocino[9',10': 17,18;10",9":25,26][1,3,6,11,14,16,19, 24]octaoxacyclohexacosino[4,5-j:13,12-j]bis[1,3]benzodioxocin, Stereoisomer **5**ONMP. Procedure C. A mixture of diol 2 (100 mg, 0.045 mmol), 50 mL of NMP, 1 g of Cs₂CO₃, and 33 mg (0.09 mmol) of 1,2-ethanediol ditosylate was stirred at 75 °C for 24 h, and a second portion of 66 mg (0.18 mmol) of 1,2-ethanediol ditosylate was added. After stirring at 75 °C for another 24 h, the solvent was removed in vacuo and the residue was dissolved in CHCl₃. The remaining solids were filtered through a 1 cm pad of Celite and the solvent was rotary evaporated, concentrated to \sim 3 mL, and poured into 100 mL of methanol. The precipitate that formed was filtered and chromatographed on a preparative TLC plate with CHCl₃ to give 47 mg (45%) of 5 \odot NMP: ¹H NMR δ -0.93 (2 H, q); -0.89 (3 H, s); -0.72 (2 H, t); 1.99 (12 H, m); 2.51 (16 H, m); 2.67 (16 H, m); 3.95 (8 H, br s); 4.06 (4 H, br s); 4.21 (4 H, d, J = 7.0 Hz); 4.42 (4 H, d, J = 7.0 Hz); 4.54 (4 H, s); 4.81 (8 H, m); 6.05 (4H, d, *J* = 7.3 Hz); 6.80 (2 H, s); 6.83 (4 H, s); 6.87 (2 H, s); 7.12 (16 H, m); 7.24 (24 H, m); FAB MS, m/e (2320.0, M⁺) 2320.2, (100). Anal. Calcd for $C_{147}H_{141}NO_{25}$: C, 76.05; H, 6.12. Found: C, 76.09; H, 6.06.

5 DMSO. Application of procedure C to 100 mg (0.045 mmol) of diol **2**, 15 mL of DMSO, 1 g of Cs₂CO₃, and 99 mg (0.27 mmol) of 1,2-ethanediol ditosylate gave 42 mg (41%) of **5 D**MSO after preparative TLC: ¹H NMR δ –0.48 (6 H, s); 2.02 (12 H, br s); 2.47 (16 H, m); 2.68 (16 H, m); 3.95 (8 H, br s); 4.04 (4 H, br s); 4.12 (4 H, d, J= 7.1 Hz); 4.28 (4 H, d, J= 7.1 Hz); 4.54 (4 H, s); 4.84 (8 H, m); 5.86 (8 H, t, J= 7.6 Hz); 6.78 (2 H, s); 6.85 (4 H, s); 6.89 (2 H, s); 7.17 (16 H, m); 7.23 (24 H, m); FAB MS, m/e (2298.9, M⁺) 2301.4 (100), 2222.8 (30). Anal. Calcd for C₁₄₄H₁₃₈O₂₅S: C, 75.18; H, 6.05. Found: C, 75.08; H, 6.05.

5 • DMA. Application of procedure C to 100 mg (0.045 mmol) of diol **2**, 20 mL of DMA, 1 g of Cs₂CO₃, and 99 mg (0.27 mmol) of 1,2-ethanediol ditosylate gave 44 mg (42%) of **5 • •** DMA after preparative TLC: ¹H NMR δ –1.67 (3 H, s); -0.56 (3 H, s); 1.64 (3 H, s); 1.96 (12 H, m); 2.48 (16 H, m); 2.67 (16 H, m); 3.92 (8 H, t, br s); 4.01 (4 H, br s); 4.14 (4 H, d, J = 7.1 Hz); 4.45 (4 H, d, J = 7.1 Hz); 4.65 (4 H, s); 4.83 (8 H, m); 5.81 (8H, t, J = 6.4 Hz); 6.75 (2 H, s); 6.83 (4 H, s); 6.89 (2 H, s); 7.16 (16 H, m); 7.22 (24 H, m); FAB MS, m/e (2308.0, M⁺), 2309.7 (100). Anal. Calcd for C₁₄₆H₁₄₁NO₂₅: C, 75.92; H, 6.15. Found: C, 75.87; H, 5.96.

5⊙**1,4-Me₂C₆H₄.** Application of procedure B to 30 mg (0.013 mmol) of **5**⊙DMA and 1 mL of *p*-xylene gave after preparative TLC with CHCl₃ 25 mg (88%) of **5**⊙1,4-Me₂C₆H₄: ¹H NMR *δ* −1.93 (6 H, s); 1.21 (4 H, m); 2.10 (8 H, m); 2.52 (16 H, m); 2.70 (16 H, m); 3.09 (4 H, t); 4.03 (4 H, br s); 4.11 (4 H, d, J = 6.9 Hz); 4.17 (4 H, d, J = 6.9 Hz); 4.18 (4 H, s); 4.21 (4 H, t); 4.86 (8 H, m); 5.69 (4H, d, J = 6.9 Hz); 5.78 (4H, d, J = 6.9 Hz); 5.97 (4 H, s); 6.82 (2 H, s); 6.85 (2 H, s); 6.94 (4 H, s); 7.17 (16 H, m); 7.23 (24 H, m); FAB MS, *m*/e (2327.0, M⁺), 2329.7 (100), 2224.0 (90). Anal. Calcd for C₁₅₀H₁₄₂O₂₄: C, 77.37; H, 6.15. Found: C, 77.68; H, 5.98.

8,9,10,11,39,40,41,42-Octahydro-1,18,26,28,53,55,63,79octaphenethyl-34,47-(epoxybutanoxy)-20,24:57,61-dimethano-2,52:17,29-dimetheno-3,51,16,30-(methynoxypropanoxymethyno)-1*H***,18***H***,26***H***,28***H***,53***H***,55***H***-bis[1,3]benzodioxocino[9,8-***d***:9',8'-***d***']bis[1,3]benzodioxocino[9',10': 17,18;10'',9'':25,26][1,3,6,11,14,16,19, 24]octaoxacyclohexacosino[4,5-***j***:13,12-***j***]bis[1,3]benzodioxocin, Stereoisomer 6 (Empty). Procedure D.** A mixture of diol **2** (100 mg (0.045 mmol), 10 mL of HMPA, 1 g of Cs₂CO₃, and 35 mg (0.09 mmol) of 1,3-propanediol ditosylate was stirred at 75 °C for 24 h, and 70 mg (0.18 mmol) more of 1,3-propanediol ditosylate was added. After stirring at 75 °C for another 24 h, the mixture was poured into 50 mL of 5% NaCl (aq). The precipitate that formed was filtered, washed with methanol, and chromatographed on a preparative TLC plate with CHCl₃ to give 46 mg (46%) of **6** (empty): ¹H NMR δ 1.99 (12 H, br s); 2.22 (2 H, m); 2.51 (16 H, m); 2.67 (16 H, m); 3.90 (12 H, m); 4.18 (4 H, t, *J* = 7.1 Hz); 4.30 (8 H, d, *J* = 7.8 Hz); 4.81 (8 H, t, *J* = 7.8 Hz); 5.78 (4 H, d, *J* = 7.3 Hz); 5.82 (4 H, d, *J* = 7.3 Hz); 6.82 (8 H, m); 7.12 (16 H, m); 7.23 (24 H, m); FAB MS, *m*/e (2234.9, M⁺), 2237.5 (100). Anal. Calcd for C₁₄₃H₁₃₄O₂₄•2H₂O: C, 75.57; H, 6.12. Found: C, 75.62; H, 6.09.

6⊙**NMP.** Application of procedure C to 100 mg (0.045 mmol) of diol **2**, 50 mL of NMP, 1 g of Cs₂CO₃, and 35 mg (0.09 mmol) of 1,3-propanediol ditosylate (first portion) and 70 mg (0.18 mmol) of 1,3-propanediol ditosylate (second portion) gave after preparative TLC with CHCl₃ 67 mg (64%) of **6**⊙NMP: ¹H NMR δ −0.90 (3 H, s); −0.82 (2 H, q); −0.60 (2 H, t); 1.98 (12 H, br s); 2.22 (2 H, m); 2.48 (16 H, m); 2.65 (16 H, m); 3.98 (12 H, m); 4.18 (4 H, t, *J* = 7.1 Hz); 4.21 (4 H, d, *J* = 7.6 Hz); 4.41 (4 H, d, *J* = 7.6 Hz); 4.85 (8 H, t, *J* = 7.8 Hz); 5.81 (8 H, t, *J* = 7.3 Hz); 6.85 (8 H, m); 7.13 (16 H, m); 7.23 (24 H, m); FAB MS, *m*/*e* (2334.0, M⁺), 2336.0 (100). Anal. Calcd for C₁₄₈H₁₄₃NO₂₅: C, 76.11; H, 6.17. Found: C, 76.17; H, 6.17.

6⊙**DMSO.** Application of procedure C to 100 mg (0.045 mmol) of diol host **2**, 50 mL of DMSO, 1 g of Cs₂CO₃, and 105 mg (0.27 mmol) of 1,3-propanediol ditosylate gave 62 mg (60%) of **6**⊙DMSO after preparative TLC with CHCl₃: ¹H NMR δ −0.51 (6 H, s); 2.03 (12 H, br s); 2.25 (2 H, m); 2.50 (16 H, m); 2.69 (16 H, m); 3.94 (12 H, m); 4.19 (4 H, t); 4.21 (8 H, t); 4.84 (8 H, t, *J* = 7.8 Hz); 5.83 (4 H, d, *J* = 7.3 Hz); 5.88 (4 H, d, *J* = 7.3 Hz); 6.84 (8 H, m); 7.15 (16 H, m); 7.24 (24 H, m); FAB MS, *m*/*e* (2312.9, M⁺), 2315.4 (100). Anal. Calcd for C₁₄₅H₁₄₀O₂₅S: C, 75.24; H, 6.10. Found: C, 75.31; H, 6.10.

6 DMA. Application of procedure C to 100 mg (0.045 mmol) of diol **2**, 50 mL of DMA, 1 g of Cs₂CO₃, and 105 mg (0.27 mmol) of 1,3-propanediol ditosylate gave 67 mg (64%) of **6 D**MA after preparative TLC with CHCl₃: ¹H NMR δ –1.68 (3 H, s); -0.53 (3 H, s); 2.01 (12 H, br s); 2.25 (2 H, m); 2.49 (16 H, m); 2.68 (16 H, m); 3.91 (12 H, m); 4.20 (8 H, d, J = 7.2 Hz); 4.39 (4 H, d, J = 7.4 Hz); 4.84 (8 H, t, J = 7.9 Hz); 5.81 (4 H, d, J = 7.5 Hz); 5.84 (4 H, d, J = 7.5 Hz); 6.82 (2 H, s); 6.84 (4 H, s); 6.87 (2 H, s); 7.15 (16 H, m); 7.24 (24 H, m); FAB MS, m/e (2322.0, M⁺), 2323.7 (100). Anal. Calcd for C₁₄₇H₁₄₃NO₂₅: C, 75.98; H, 6.20. Found: C, 76.16; H, 6.03.

6 \odot **1**,**4**-**Me**₂**C**₆**H**₄. Application of procedure B to 30 mg (0.013 mmol) of **6** (empty) and 1 mL of *p*-xylene gave after preparative TLC with CHCl₃ 27 mg (90%) of **6** \odot 1,4-Me₂C₆H₄: ¹H NMR δ -2.01 (6 H, s); 1.67 (4 H, br s); 1.97 (2 H, m); 2.09 (8 H, br s); 2.52 (16 H, m); 2.70 (16 H, m); 3.36 (4 H, t, *J* = 6.9 Hz); 3.66 (4 H, br s); 4.12 (8 H, hidden); 4.12 (4 H, d, *J* = 6.9 Hz); 4.87 (8 H, t, *J* = 7.8 Hz); 5.71 (8 H, dd, *J* = 7.0 Hz); 5.94 (4H, s); 6.85 (4 H, s); 6.94 (4 H, s); 7.16 (16 H, m); 7.23 (24 H, m); FAB MS, *m*/*e* (2341.0, M⁺), 2342 (100), 2236 (50). Anal. Calcd for C₁₅₁H₁₄₄O₂₄: C, 77.41; H, 6.20. Found: C, 77.29; H, 6.12.

8,9,10,11,40,41,42,43-Octahydro-1,18,26,28,54,56,64,81octaphenethyl-34,48-(epoxybutanoxy)-20,24:58,62-dimethano-2,53:17,29-dimetheno-3,52,16,30-(methynoxybutanoxymethyno)-1H,18H,26H,28H,39H,54H,56H-bis[1,3]benzodioxocino[9,8-d:9',8'-d']bis[1,3]benzodioxocino-[9',10':4,5;10",9":12,13][1,3,6,11,14,16,19,25]octaoxacycloheptacosino[17,18-j:27,26-j]bis[1,3]benzodioxocin, Stereoisomer 80CHCl₃. A mixture of 100 mg (0.045 mmol) of diol 2, 50 mL of NMP, 1 g of Cs₂CO₃, and 36 mg (0.09 mmol) of 1,5-pentanediol dimesylate was stirred at 25 °C for 5 h, and then the temperature was raised to 50 °C and the mixture stirred for 24 h. The solution was stirred for another 24 h after the addition of 36 mg (0.09 mmol) of 1,5-pentanediol dimesylate. The product was isolated as in procedure A to give after preparative TLC with CHCl₃ 81 mg (79%) of 8⊙CHCl₃: ¹H NMR δ 1.82 (6 H, m); 1.97 (12 H, br s); 2.49 (16 H, m); 2.68 (16 H, m); 3.88 (4 H, m); 3.97 (12 H, br s); 4.20 (4 H, d, J = 7.0 Hz); 4.23 (4 H, d, J = 7.0 Hz); 4.83 (8 H, m); 5.81 (4 H, d, J = 6.9 Hz); 5.86 (4 H, d, J = 6.9 Hz); 6.80 (2 H, s);6.82 (2 H, s); 6.84 (4 H, s); 7.17 (16 H, m); 7.23 (24 H, m); FAB

MS, m/e (2380.9, M⁺), 2384.0 (100), 2264.6 (98). Anal. Calcd for $C_{146}H_{139}Cl_3O_{24}$: C, 73.56; H, 5.88. Found: C, 73.58; H, 5.60.

8 CHCl₂CHCl₂. Application of procedure B to 30 mg (0.013 mmol) of **8 C**HCl₃ and 1 mL of tetrachloroethane gave after preparative TLC with CHCl₃ 28 mg (92%) of **8 C**HCl₂-CHCl₂: ¹H NMR δ 1.80 (6 H, m); 1.97 (12 H, br s); 2.49 (16 H, m); 2.69 (16 H, m); 3.88 (4 H, m); 3.98 (12 H, br s); 4.29 (2 H, s); 4.35 (8 H, dd); 4.82 (8 H, m); 5.78 (4 H, d, J = 7.1 Hz); 5.83 (4 H, d, J = 7.1 Hz); 6.79 (2 H, s); 6.82 (2 H, s); 6.84 (4 H, s); 7.17 (16 H, m); 7.22 (24 H, m); FAB MS, m/e (2428.8, M⁺), 2431 (100), 2263 (60). Anal. Calcd for C₁₄₇H₁₄₀Cl₄O₂₄3H₂O: C, 71.01; H, 5.92. Found: C, 70.74; H, 5.74.

8©1,4-(**MeO**)₂C₆H₄. **Procedure F.** Into a Pyrex test tube capped with a rubber septum were placed 30 mg (0.013 mmol) of **8**©CHCl₃, 180 mg of 1,4-dimethoxybenzene (1.3 mmol), and 1 mL of Ph₂O. This mixture was heated at 165 °C for 3 d and poured into 50 mL of methanol. The precipitate that formed was filtered (without further purification) to give 28 mg (90%) of **8**©1,4-(MeO)₂C₆H₄: ¹H NMR δ –0.41 (6 H, s); 1.56 (6 H, br s); 1.81 (8 H, br s); 2.03 (4 H, br s); 2.50 (16 H, m); 2.69 (16 H, m); 3.58 (4 H, m); 3.62 (4 H, m); 4.04 (4 H, br s); 4.08 (4 H, br s); 4.28 (8 H, t, *J* = 7.1 Hz); 4.90 (8 H, t, *J* = 7.7 Hz); 5.74 (8 H, t, *J* = 7.0 Hz); 5.84 (4 H); 6.82 (4 H, s); 6.85 (2 H, s); 6.86 (2 H, s); 7.17 (16 H, m); 7.24 (24 H, m); FAB MS, *m/e* (2401.0, M⁺), 2402.6 (100). Anal. Calcd for C₁₅₃H₁₄₈O₂₆H₂O: C, 75.91; H, 6.25. Found: C, 76.10; H, 5.90.

8⊙1,3-(MeO)₂C₆H₄. Procedure G. Into a Pyrex test tube capped with a rubber septum were placed 30 mg (0.013 mmol) of **8**⊙CHCl₃ and 1 mL of 1,3-dimethoxybenzene. This mixture was heated at 165 °C for 3 d and poured into 50 mL of methanol. The precipitate that formed was filtered and chromatographed on a preparative TLC plate with CHCl₃ to give 28 mg (89%) of **8**⊙1,3-(MeO)₂C₆H₄: ¹H NMR δ −0.55 (6 H, s); 1.53 (6 H, br s); 1.87 (8 H, br s); 2.03 (4 H, br s); 2.50 (16 H, m); 3.55 (4 H, m); 3.60 (4 H, m); 4.09 (8 H, br s); 4.33 (8 H, t, *J* = 6.3 Hz); 4.92 (8 H, m); 5.02 (2 H, d); 5.48 (1 H, s); 5.74 (8 H, th); 7.24 (24 H, m); FAB MS, *m/e* (2401.0, M⁺), 2402.6 (100). Anal. Calcd for C₁₅₃H₁₄₈O₂₆: C, 76.48; H, 6.21. Found: C, 76.31; H, 6.05.

8 \odot **1,2**-(**MeO**)₂**C**₆**H**₄. Application of procedure G to 8 \circ CHCl₃ (30 mg, 0.013 mmol) and 1 mL of 1,2-(MeO)₂C₆H₄ gave, after preparative TLC with CHCl₃, 26 mg (86%) of **8** \odot 1,2-(MeO)₂C₆H₄: ¹H NMR δ 1.62 (8 H, br s); 1.78 (6 H, br s); 2.04 (4 H, br s); 2.50 (16 H, m); 2.68 (16 H, m); 3.60 (4 H, br s); 3.71 (4 H, m); 4.05 (8 H, br s); 4.34 (8 H, t); 4.83 (8 H, m); 5.46 (2 H, br s); 5.70 (8 H, t, *J* = 7.0 Hz); 6.87 (4 H, s); 6.91 (2 H, s); 6.93 (2 H, s); 7.17 (16 H, m); 7.23 (24 H, m); FAB MS *m*/*e* (2401.0, M⁺), 2401.1 (100). Anal. Calcd for C₁₅₃H₁₄₈O₂₆: C, 76.48; H, 6.21. Found: C, 76.31; H, 6.05.

8 • Naphthalene. Application of procedure G to 30 mg (0.013 mmol) of **8** • CHCl₃, 166 mg of naphthalene (1.3 mmol), and 1.5 mL of Ph₂O gave, after preparative TLC with CHCl₃, 26 mg (84%) of **8** • naphthalene: ¹H NMR δ 1.31 (8 H, m); 1.85–2.10 (6 H, m); 2.21 (4 H, br s); 2.55 (16 H, m); 2.71 (16 H, m); 3.15 (4 H, m); 3.21 (4 H, m); 3.36 (4 H, br s); 4.21 (4 H, br s); 5.62 (8 H, dd, J= 7.0 Hz); 6.93 (4 H, s); 7.09 (2 H, s); 7.11 (2 H, s); 7.17 (16 H, m); 7.24 (24 H, m); FAB MS *m/e* (2391.0, M⁺), 2391.8 (100), 2265.5, (80). Anal. Calcd for C₁₅₅H₁₄₆O₂₄: C, 77.80; H, 6.15. Found: C, 78.10; H, 6.29.

8⊙4-**MeC**₆**H**₄**OMe.** Application of procedure G to 30 mg (0.013 mmol) of **8**⊙CHCl₃ and 1 mL of 4-MeC₆H₄OMe gave 29 mg (92%) of **8**⊙4-MeC₆H₄OMe: ¹H NMR δ −1.98 (3 H, s); −0.28 (3 H, s); 1.63 (8 H, br s); 1.84 (6 H, br s); 2.04 (4 H, br s); 2.52 (16 H, m); 2.69 (16 H, m); 3.46 (2 H, m); 3.59 (2 H, m); 3.70 (4 H, br s); 4.04 (8 H, br s); 4.14 (2 H, d, J = 7.0 Hz); 4.16 (2 H, d, J = 7.0 Hz); 4.22 (2 H, d, J = 7.0 Hz); 5.71 (2 H, d, J = 6.9 Hz); 5.73 (2 H, d, J = 6.9 Hz); 5.71 (2 H, d, J = 6.9 Hz); 5.73 (2 H, d, J = 6.9 Hz); 5.78 (2 H, d, J = 8.1 Hz); 6.87 (2 H, d, J = 4.0 Hz); 6.92 (2 H, d, J = 4.0 Hz); 7.17 (16 H, m); 7.24 (24 H, m); FAB MS m/e (2385.0, M⁺), 2386.3 (100), 2265.5 (40). Anal. Calcd for C₁₅₃H₁₄₈O₂₅: C, 76.99; H, 6.25. Found: C, 77.16; H, 6.24.

8 \odot **2**-**HOC**₆**H**₄**Me.** Application of procedure G to 30 mg (0.013 mmol) of **8** \odot CHCl₃ and 1 mL of *o*-cresol gave, after preparative TLC with CHCl₃, 28 mg (90%) of **8** \odot 2-HOC₆H₄-Me: ¹H NMR δ –1.68 (3 H, s); 1.68–2.04 (18 H, m); 2.51 (16 H, m); 2.69 (16 H, m); 3.22 (1 H, t); 3.68–4.10 (16 H, m); 4.15 (2 H, d, *J* = 7.1 Hz); 4.29 (4 H, d, *J* = 7.7 Hz); 4.36 (2 H, d, *J* = 7.1 Hz); 5.78 (2 H, d, *J* = 7.0 Hz); 5.83 (2 H, d, *J* = 7.1 Hz); 5.78 (2 H, d, *J* = 7.0 Hz); 5.83 (2 H, d, *J* = 7.0 Hz); 5.98 (2 H, d); 6.14 (2 H, d); 6.21 (2 H, s); 6.71–7.08 (8 H, m); 7.17 (16 H, m); 7.23 (24 H, m); FAB MS m/e (2371.0, M⁺) 2372, (100), 2265 (80). Anal. Calcd for C₁₅₂H₁₄₆O₂₅: C, 76.94; H, 6.20. Found: C, 76.98; H, 6.23.

6,34,35,36,37,65-Hexahydro-13,21,23,48,50,76,84-octaphenethyl-29,42-(epoxybutanoxy)-15,19:52,56-dimethano-12,24:47,59-dimetheno-11,25,46,60-(methynoxybutanoxymethyno)-13H,21H,23H,48H,50H,58H-bis[1,3]benzodioxocino[9',10':4,5;10'',9'':12,13]bis[1,3]benzodioxocino[9",8":4',5'][1,3]benzodioxocino[9',10': 17,18;10",9":25,26][1,3,6,11,14,16,19, 24]octaoxacyclohexacosino[8,9-b]quinoxaline, Stereoisomer 90NMP. Procedure E. A mixture of 100 mg (0.045 mmol) of diol 2, 50 mL of NMP, 1 g of Cs₂CO₃, and 20 mg (0.09 mmol) of 2,3-bis-(bromomethyl)quinoxaline was stirred at 25 °C for 5 h, and then the temperature was raised to 50 °C and the mixture was stirred for 24 h. The solution was stirred for another 24 h after the addition of 20 mg (0.09 mmol) of 2,3-bis(bromomethyl)quinoxaline. The remaining solids were filtered through a 1 cm pad of Celite, and the solvent was rotary evaporated and concentrated to ~ 3 mL and poured into 100 mL of methanol. The precipitate that formed was filtered and chromatographed on a preparative TLC plate with CHCl₃ to give 86 mg (78%) of $9 \odot NMP$: ¹H NMR $\delta - 1.17$ (3 H, s); -1.15 (2 H, q); -0.96 (2 H, t); 1.80 (2 H, t); 1.97 (12 H, m); 2.47 (16 H, m); 2.66 (16 H, m); 3.92 (12 H, br s); 4.14 (4 H, d, J = 7.3Hz); 4.38 (4 H, d, J = 7.3 Hz); 4.71 (4 H, t, J = 7.7 Hz); 4.82 (4 H, t, J = 7.7 Hz); 5.44 (4 H, s); 5.76 (8 H, d, J = 7.2 Hz); 6.81 (4 H, s); 6.84 (2 H, s); 6.93 (2 H, s); 7.15 (16 H, m); 7.23 (24 H, m); 7.77 (2 H, m); 8.08 (2 H, m); FAB MS, m/e (2448.0, M⁺), 2450.7 (100), 2351.0 (60). Anal. Calcd for $C_{155}H_{145}$ -N₃O₂₅: C, 75.99; H, 5.97; N, 1.72. Found: C, 76.09; H, 6.13; N. 1.63.

9©**DMSO.** Application of procedure E to 100 mg (0.045 mmol) of diol **2**, 50 mL of DMSO, 1 g of Cs_2CO_3 , and 40 mg (0.18 mmol) of 2,3-bis(bromomethyl)quinoxaline gave 79 mg (72%) of **9**©DMSO after preparative TLC (CHCl₃): ¹H NMR δ –0.71 (6 H, s); 2.03 (12 H, m); 2.46 (16 H, m); 2.67 (16 H, m); 3.86 (12 H, m); 4.11 (4 H, d, J = 7.4 Hz); 4.15 (4 H, d, J = 7.4 Hz); 4.50 (4 H, t, J = 7.6 Hz); 5.82 (4 H, t, J = 7.6 Hz); 5.43 (4 H, s); 5.78 (4 H, d, J = 7.5 Hz); 5.82 (4 H, d, J = 7.5 Hz); 5.43 (4 H, s); 6.85 (2 H, s); 6.91 (2 H, s); 7.14 (16 H, m); 7.24 (24 H, m); 7.79 (2 H, m); 8.13 (2 H, m); FAB MS, m/e (2427.0, M⁺), 2430.1 (100), 2351.2 (50). Anal. Calcd for $C_{152}H_{142}N_2O_{25}S$: C, 75.17; H, 5.89. Found: C, 74.95; H, 6.13.

901,4-(MeO)₂C₆H₄. Application of procedure D to 100 mg (0.045 mmol) of diol 2, 10 mL of HMPA, 1 g of Cs₂CO₃, 620 mg (4.5 mmol) of 1,4-dimethoxybenzene, 20 mg (0.09 mmol) of 2,3-bis(bromomethyl)quinoxaline, and an additional 20 mg (0.09 mmol) of 2,3-bis(bromomethyl)quinoxaline (maximum temperature 50 °C) gave after preparative TLC with CHCl₃ 38 mg (34%) of $9 \odot 1, 4$ -(MeO)₂C₆H₄: ¹H NMR δ -0.45 (6 H, s); 1.08 (4 H, m); 2.05 (8 H, m); 2.51 (16 H, m); 2.71 (16 H, m); 3.12 (4 H, m); 4.01 (4 H, m); 4.05 (4 H, d, J = 7.0 Hz); 4.21 (4 H, m); 4.39 (4 H, d, J = 7.0 Hz); 4.78 (4 H, t, J = 7.7 Hz); 4.93 (4 H, t, J = 7.7 Hz); 5.24 (4 H, s); 5.59 (4 H, d, J = 6.9 Hz);5.76 (4 H, d, J = 6.9 Hz); 5.81 (4 H, s); 6.81 (2 H, s); 6.84 (4 H, s); 6.88 (2 H, s); 7.17 (16 H, m); 7.23 (24 H, m); 7.73 (2 H, m); 7.96 (2 H, m); FAB MS, *m/e* (2487.0, M⁺), 2488.9 (100). Anal. Calcd for C₁₅₈H₁₄₆N₂O₂₆: C, 76.25; H, 5.91. Found: C, 76.07; H. 6.05.

61,62,63,64-Tetrahydro-6,14,16,44,46,54,75,84-octaphenethyl-33*H*-22,38-(epoxybutanoxy)-8,12:48,52-dimethano-5,17:28,32:43,55-trimetheno-4,18,42,56-(methynoxybutanoxymethyno)-6*H*,14*H*,16*H*,27*H*,42*H*,46*H*,54*H*-bis[1,3]benzodioxocino[9,8-*d*:9',8'-*d*]bis[1,3]benzodioxocino-[9',10':4,5;10'',9'':12,13][1,3,6,11,14,16,19,27]octaoxacyclononacosino[17,18-*j*:29,28-*j*]bis[1,3]benzodioxocin, Stereoiso**mer 10 •CHCl**₃. Application of procedure E to 100 mg (0.045 mmol) of diol 2, 50 mL of NMP, 1 g of Cs₂CO₃, 16 mg (0.09 mmol) of 1,3-bis(chloromethyl)benzene, and 16 mg (0.09 mmol) of additional dichloride gave after preparative TLC with CHCl₃: 88 mg (81%) of **10 •C**HCl₃: ¹H NMR δ 1.94 (12 H, m); 2.48 (16 H, m); 2.68 (16 H, m); 3.86 (4 H, br s); 3.96 (4 H, br s); 4.04 (4 H, br s); 4.24 (8 H, t, *J* = 7.5 Hz); 4.85 (8 H, t, *J* = 7.8 Hz); 5.06 (4 H, s); 5.73 (4 H, d, *J* = 7.1 Hz); 5.89 (4 H, d, *J* = 7.1 Hz); 6.82 (2 H, s); 6.87 (6 H, s); 7.03 (2 H, d); 7.17 (16 H, m); 7.23 (24 H, m); 7.74 (1 H, s); FAB MS m/e (2414.9, M⁺) 2419 (40), 2299 (100). Anal. Calcd for C₁₄₉H₁₃₇Cl₃O₂₄: C, 74.01; H, 5.71. Found: C, 73.76; H, 5.78.

10 \odot **1,4-(MeO)**₂**C**₆**H**₄. Application of procedure G to 30 mg (0.013 mmol) of **10** \odot CHCl₃, 180 mg of 1,4-dimethoxybenzene (1.3 mmol), and 1 mL of Ph₂O gave 28 mg (90%) of **10** \odot 1,4-(MeO)₂C₆H₄: ¹H NMR δ –0.38 (6 H, s); 1.49 (8 H, br s); 2.05 (4 H, br s); 2.53 (16 H, m); 2.71 (16 H, m); 3.55 (4 H, br s); 3.58 (4 H, br s); 4.09 (4 H, br s); 4.25 (4 H, d, J = 7.0 Hz); 4.27 (4 H, d, J = 7.0 Hz); 4.91 (8 H, t); 5.10 (4 H, s); 5.68 (4 H, d, J = 6.9 Hz); 5.73 (4 H, d, J = 6.9 Hz); 5.84 (4 H, s); 6.88 (2 H, s); 6.91 (2 H, s); 7.19 (16 H, m); 7.24 (24 H, m); 7.32 (2 H, d, J = 7.6 Hz); 7.69 (1 H, s); FAB MS *m/e* (2435.0, M⁺) 2435 (100). Anal. Calcd for C₁₅₆H₁₄₆O₂₆: C, 76.89; H, 6.04. Found: C, 76.98; H, 6.00.

10⊙**1,3**-(**MeO**)₂**C**₆**H**₄. Application of procedure F to 30 mg (0.013 mmol) of **10**⊙CHCl₃ and 1 mL of 1,3-dimethoxybenzene gave after preparative TLC with CHCl₃, 27 mg (89%) of **10**⊙1,3-(MeO)₂C₆H₄: ¹H NMR δ −0.53 (6 H, s); 1.45 (8 H, br s); 2.05 (4 H, br s); 2.53 (16 H); 2.72 (16 H, m); 3.49 (4 H, br s); 3.55 (4 H, br s); 4.08 (4 H, br s); 4.29 (8 H, d, *J* = 6.8 Hz); 4.91 (8 H, t); 5.03 (2 H, d); 5.13 (4 H, s); 5.48 (1 H, s); 5.66 (4 H, d, *J* = 7.0 Hz); 5.73 (4 H, d, *J* = 6.9 Hz); 6.83 (4 H, s); 6.89 (2 H, s); 6.93 (2 H, s); 7.19 (16 H, m); 7.24 (24 H, m); 7.43 (1 H, t); 7.78 (1 H, s); FAB MS *m*/*e* (2435.0, M⁺) 2436 (100). Anal. Calcd for C₁₅₆H₁₄₆O₂₆: C, 76.89; H, 6.04. Found: C, 76.66; H, 6.23.

10 \odot **1,2**-(**MeO**)₂**C**₆**H**₄. Application of procedure F to 30 mg (0.013 mmol) of **10** \odot CHCl₃ and 1 mL of 1,2-(MeO)₂C₆H₄ gave after preparative TLC with CHCl₃, 27 mg (87%) of **10** \odot 1,2-(MeO)₂C₆H₄: ¹H NMR δ 1.59 (8 H, br s); 2.03 (4 H, br s); 2.50 (16 H, m); 2.68 (16 H, m); 3.58 (4 H, br s); 3.69 (4 H, m); 4.08 (4 H, br s); 4.30 (8 H, t, *J* = 7.5 Hz); 4.88 (8 H, m); 5.12 (4 H, s); 5.45 (2 H, br s); 5.71 (8 H, d, *J* = 7.8 Hz); 6.89 (4 H, s); 6.99 (2 H, s); 7.01 (2 H, s); 7.18 (16 H, m); 7.24 (24 H, m); 7.34 (2 H, d); 7.51 (1 H, t); 7.71 (1 H, s); FAB MS *m*/*e* (2435.0, M⁺) 2436 (100). Anal. Calcd for C₁₅₆H₁₄₆O₂₆: C, 76.89; H, 6.04. Found: C, 76.62; H, 6.09.

10 Naphthalene. Application of procedure G to 30 mg (0.013 mmol) of **10 CHCl**₃, 166 mg of naphthalene (1.3 mmol), and 1.5 mL of Ph₂O gave after preparative TLC with CHCl₃ 27 mg (85%) of **10 O** naphthalene: ¹H NMR δ 1.31 (8 H, m); 2.20 (4 H, m); 2.56 (16 H, m); 2.71 (16 H, m); 3.12 (4 H, m); 3.21 (4 H, m); 3.35 (4 H, m); 4.19 (4 H, br s); 4.30 (4 H, d, J = 7.1 Hz); 4.34 (4 H, d, J = 7.1 Hz); 4.89 (8 H, m); 5.21 (4 H, s); 5.54 (4 H, d, J = 6.9 Hz); 5.61 (4 H, d, J = 6.9 Hz); 6.98 (4 H, s); 7.12 (2 H, s); 7.18 (16 H, m); 7.24 (24 H, m); 7.41 (1 H, t); 8.08 (1 H, s); FAB MS m/e (2425.0, M⁺) 2425.1 (100). Anal. Calcd for C₁₅₈H₁₄₄O₂₄·2H₂O: C, 77.05; H, 6.06. Found: C, 77.04; H, 5.95.

10 \odot **4**-**MeC**₆**H**₄**OMe.** Application of procedure F to 30 mg (0.013 mmol) of **10** \odot CHCl₃ and 1 mL of 4-MeC₆H₄OMe gave 28 mg (90%) of **10** \odot 4-MeC₆H₄OMe: ¹H NMR δ –1.96 (3 H, s); -0.26 (3 H, s); 1.58 (8 H, br s); 2.03 (4 H, br s); 2.50 (16 H, m); 2.69 (16 H, m); 3.39 (2 H, m); 3.58 (2 H, m); 3.62 (2 H, m); 3.71 (2 H, m); 4.02 (4 H, br s); 4.16 (2 H, d, *J* = 7.0 Hz); 4.18

(2 H, d, J = 7.0 Hz); 4.22 (2 H, d, J = 7.0 Hz); 4.29 (2 H, d, J = 7.0 Hz); 4.89 (8 H, m); 5.62 (4 H, d, J = 8.4 Hz); 5.62 (4 H, t, J = 6.8 Hz); 5.73 (2 H, d, J = 6.9 Hz); 5.80 (2 H, d, J = 6.9 Hz); 5.87 (2 H, d, J = 8.5 Hz); 6.01 (2 H, d, J = 8.5 Hz); 6.80–7.01 (8 H, m); 7.17 (16 H, m); 7.24 (24 H, m); 7.38 (1 H, t); 7.80 (1 H, s); FAB MS m/e (2419.0, M⁺) 2419.9 (100). Anal. Calcd for C₁₅₆H₁₄₆O₂₅: C, 77.40; H, 6.08. Found: C, 77.71; H, 6.26.

10 \odot **1,2,3**-(**MeO**)₃C₆H₃. Application of procedure F to 30 mg (0.013 mmol) of **10** \odot CHCl₃, 220 mg of 1,2,3-trimethoxybenzene (1.3 mmol), and 1 mL of Ph₂O gave after preparative TLC with CHCl₃ 26 mg (80%) of **10** \odot 1,2,3-(MeO)₃C₆H₃: ¹H NMR δ –0.41 (6 H, s); 1.45 (8 H, br s); 2.08 (4 H, br s); 2.51 (16 H, m); 2.71 (16 H, m); 3.05 (3 H, s); 3.62 (8 H, br s); 4.12 (4 H, br s); 4.39 (4 H, d, J = 7.2 Hz); 4.49 (4 H, d, J = 7.2 Hz); 4.92 (8 H, m); 5.18 (4 H, s); 5.67 (4 H, d, J = 6.9 Hz); 5.73 (4 H, d, J = 6.9 Hz); 6.86 (4 H, s); 6.90 (2 H, s); 6.98 (2 H, s); 7.18 (16 H, m); 7.24 (24 H, m); 7.34 (2 H, d); 7.46 (1 H, t); 7.60 (1 H, s); FAB MS m/e (2465.0, M⁺) 2468.3 (100). Anal. Calcd for C₁₅₇H₁₄₈O₂₇: C, 76.44; H, 6.05. Found: C, 76.26; H, 5.94.

61,62,63,64-Tetrahydro-6,14,16,44,46,54,75,84-octaphenethyl-33H-22,38-(epoxybutanoxy)-8,12:48,52-dimethano-5,17:43,55-dimetheno-4,18,42,56-(methynoxybutanoxymethyno)-28,32-nitrilo-6H,14H,16H,27H,44H,46H,54H-bis-[1,3]benzodioxocino[9,8-d:9',8'-d']bis[1,3]benzodioxocino[9',10':4,5;10",9":12,13][1,3,6,11,14,16,19,27]octaoxacyclononacosino[17,18-j:29,28-j']bis[1,3]benzodioxocin, Stereoisomer 11OCHCl₃. A mixture of 100 mg (0.045 mmol) of diol 2, 50 mL of NMP, 1 g of Cs₂CO₃, and 16 mg (0.09 mmol) of 2,6-bis(chloromethyl)pyridine was stirred at 25 °C for 5 h, and then the temperature was raised to 50 °C and the mixture stirred for 24 h. The solution was stirred for another 24 h after the addition of 16 mg (0.09 mmol) of 2,6-bis-(chloromethyl)pyridine. The solvent was removed in vacuo, and the residue was dissolved in CHCl₃. The remaining solids were filtered through a 1 cm pad of Celite, and the filtrate was rotary evaporated, concentrated to \sim 3 mL, and poured into 100 mL of methanol. The precipitate that formed was filtered and chromatographed on a preparative TLC plate with CHCl₃ to give 84 mg (77%) of 11 OCHCl₃: ¹H NMR δ 1.91 (8 H, m); 1.98 (4 H, m); 2.49 (16 H, m); 2.69 (16 H, m); 3.87 (4 H, m); 3.90 (4 H, m); 3.98 (4 H, br s); 4.20 (4 H, d); 4.32 (4 H, d); 4.83 (8 H, m); 5.12 (4 H, s); 5.62 (4 H, d, J = 6.2 Hz); 5.82 (4 H, d, J = 6.2 Hz); 6.81 (2 H, s); 6.83 (4 H, s); 6.88 (2 H, s); 7.03 (2 H, d); 7.17 (16 H, m); 7.24 (24 H, m); 7.35 (1 H, br s); FAB MS m/e (2415.9, M⁺) 2420 (100), 2300 (25). Anal. Calcd for C₁₄₈H₁₃₆Cl₃NO₂₄: C, 73.48; H, 5.78. Found: C, 73.22; H, 5.57.

11 ONaphthalene. Application of procedure F to 30 mg (0.013 mmol) of **11 OCHCl**₃, 166 mg of naphthalene (1.3 mmol), and 1.5 mL of Ph₂O gave after preparative TLC with CHCl₃ 27 mg (87%) of **11 Onaphthalene**: ¹H NMR δ 1.20 (8 H, m); 2.19 (4 H, m); 2.57 (16 H, m); 2.71 (16 H, m); 3.12 (8 H, m); 3.37 (4 H, m); 4.19 (4 H, br s); 4.20 (4 H, d, J = 7.0 Hz); 4.30 (4 H, d, J = 7.0 Hz); 4.87 (8 H, m); 5.38 (4 H, s); 5.53 (4 H, d, J = 6.9 Hz); 5.58 (4 H, d, J = 6.9 Hz); 6.94 (4 H, s); 7.12 (2 H, s); 7.18 (16 H, m); 7.24 (24 H, m); 7.64 (1 H, t); FAB MS m/e (2426.0, M⁺) 2427.9 (100), 2300.5 (80). Anal. Calcd for C₁₅₇H₁₄₃NO₂₄: C, 77.67; H, 5.94. Found: C, 77.29; H, 5.83.

11 \odot **1,2-(MeO)**₂**C**₆**H**₄. Application of procedure F to 30 mg (0.013 mmol) of **11** \odot CHCl₃ and 1 mL of 1,2-(MeO)₂C₆H₄ gave after preparative TLC with CHCl₃ 27 mg (84%) of **11** \odot 1,2-(MeO)₂C₆H₄: ¹H NMR δ 1.58 (8 H, br s); 2.05 (4 H, br s); 2.53 (16 H, m); 2.70 (16 H, m); 3.57 (4 H, br s); 3.67 (4 H, m); 4.11 (4 H, br s); 4.25 (8 H, m); 4.87 (8 H, m); 5.27 (4 H, s); 5.55 (2 H, br s); 5.69 (8 H, d, *J* = 7.0 Hz); 6.89 (4 H, s); 6.97 (2 H, s); 6.98 (2 H, s); 7.18 (16 H, m); 7.24 (24 H, m); 7.34 (2 H, d); 7.59 (1 H, br s); FAB MS *m*/*e* (2436.0, M⁺) 2438.6 (100), 2300.9 (30). Anal. Calcd for C₁₅₅H₁₄₅NO₂₆: C, 76.37; H, 6.00. Found: C, 76.53; H, 5.99.

11 \odot **1,3-(MeO)**₂**C**₆**H**₄. Application of procedure F to 30 mg (0.013 mmol) of **11** \odot CHCl₃ and 1 mL of 1,3-dimethoxybenzene gave after preparative TLC with CHCl₃, 27 mg (86%) of **11** \odot 1,3-(MeO)₂C₆H₄: ¹H NMR δ –0.50 (6 H, s); 1.40 (8 H, m); 2.08 (4 H, br s); 2.52 (16 H, m); 2.71 (16 H, m); 3.53 (8 H, m); 4.11 (4 H, br s); 4.35 (8 H, d, J = 6.8 Hz); 4.90 (8 H, t); 5.00 (2 H, d); 5.34 (4 H, s); 5.49 (1 H, s); 5.70 (8 H, d, J = 7.0 Hz);

6.83 (4 H, s); 6.89 (2 H, s); 6.93 (2 H, s); 7.19 (16 H, m); 7.24 (24 H, m); 7.74 (1 H, br s); FAB MS m/e (2436.0, M⁺) 2437.9 (90), 2300.4 (100). Anal. Calcd for $C_{155}H_{145}NO_{26}$: C, 76.37; H, 6.00. Found: C, 76.40; H, 5.98.

11 \odot **4-MeC₆H₄OMe.** Application of procedure F to 30 mg (0.013 mmol) of **11** \odot CHCl₃ and 1 mL of 4-MeC₆H₄OMe gave 28 mg (90%) of **11** \odot 4-MeC₆H₄OMe: ¹H NMR δ –1.98 (3 H, s, guest CH₃); –0.29 (3 H, s); 1.58 (8 H, br s); 2.02 (4 H, br s); 2.50 (16 H, m); 2.69 (16 H, m); 3.38 (2 H, m); 3.54 (2 H, m); 3.61 (2 H, m); 3.68 (2 H, m); 4.02 (4 H, br s); 4.15 (8 H, m); 4.87 (8 H, m); 5.48 (4 H, d); 5.72 (8 H, m); 5.78 (2 H, d); 5.90 (2 H, d); 6.80–7.01 (8 H, m); 7.17 (16 H, m); 7.24 (24 H, m); 7.71 (1 H, m); FAB MS *m*/*e* (2420.0 M⁺) 2421.2 (100), 2300.7 (30). Anal. Calcd for C₁₅₅H₁₄₅NO₂₅: C, 76.87; H, 6.03. Found: C, 77.13; H, 6.00.

Kinetics of Decomplexations of $7 \odot$ guest, $8 \odot$ guest, and $10 \odot$ guest in CDCl₃. Solutions of 5-7 mg of the three

hemicarceplexes were dissolved in 0.5 mL of $CDCl_3$ at 25 °C, and changes in guest-proton spectra were monitored over the periods indicated in Table 3. The half-lives for decomplexation were calculated from the first-order rate constants for either disappearance of complexed guest and/or appearance of free guest.

Supporting Information Available: Details of crystallographic data collection and refinement for **8** \odot 4-MeC₆H₄-OMe \cup 2C₆H₅NO₂ and **10** \odot CHCl₃ \cup 2C₆H₅NO₂ 2C₆H₅NO₂ (2 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

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